Analytical method development and validation for iron and ascorbic acid in ferrous bisglycinate effervescent tablets by colorimetry and reverse Phase-High-performance liquid chromatography

A. Srivalli1, Rameshwar Nalawade2, B. S. Muddukrishna1, Gude Sai Sushmitha1, S. G. Vasantharaju1

1Department of Pharmaceutical Quality Assurance, Manipal College of Pharmaceutical Sciences, Manipal University, Manipal, Karnataka, India, 2Analytical Development Center, Steer Life India Ltd., Bengaluru, Karnataka, India

ABSTRACT

Introduction: The new ferrous bisglycinate effervescent formulation is a combination of iron and ascorbic acid where iron is in complex form with glycine. No analytical method was reported so far for the analysis of iron and ascorbic acid in effervescent tablets. Hence, an attempt was made to develop and validate the colorimetric and RP-HPLC method. Objective: To develop and validate a colorimetric and RP-HPLC analytical method for ascorbic acid and iron by optimizing the concentration of sulphuric acid and volume of sodium acetate to breakdown ferrous bisglycinate complex. Methods: Colorimetric method for estimation of total iron: The wavelength used was 510 nm with diluent-1 as 0.5 M sulfuric acid and 1.2 M sodium acetate 5 mL. The diluent-2 was Milli Q-water. 10 mL of 1,10 phenanthroline (1 mg/mL) used as complexing agent. Analytical method development for ascorbic acid using HPLC: A reversed phase-high-performance liquid chromatography method was developed for ascorbic acid employing Inertsil octadecylsilyl column (250 mm ×4.6 mm; 5 μm) 20 mM potassium dihydrogen phosphate (pH - 2.5) and methanol (97:03) was used as mobile phase. An isocratic program was followed using 0.3 M hydrochloric acid with 5% ortho phosphoric acid as diluents with 1 ml/min flowrate. Results: The validated data of colorimetric method showed that the method was linear over a range of 0.625-3.75 μg/mL (r² = 0.999) , accurate and precise. Validated data showed that the method was linear over a range of 4.0-60.0 μg/mL (r² = 0.999). The accuracy of the method ranged from 100% to 102%. The relative standard deviation value for method precision was 0.8%. Conclusion: The proposed method is simple and more economical in comparison with other reported methods discussed in literature and it has all the potential for routine quality control analysis.

INTRODUCTION

Iron is found to be fourth greatest abundant element in the earth’s crust. In general, an average adult human body contains 4-6 g of iron. In human beings, majorly iron is present in a blood protein called hemoglobin. When the dietary intake of iron is deficient it results in a condition called anemia [1].

Iron deficiency anemia is a worldwide health problem. The prevalence of anemia is very high in India. The World Health Organization estimated that about two billion people are anemic as their hemoglobin concentrations are below recommended thresholds. The major causes of anemia are dietary iron deficiency, infectious diseases which decrease amount of red blood cells (RBCs), deficiencies of other key micronutrients including folic acid, vitamin B12, and retinol or inherited conditions that affect RBCs such as thalassemia. Iron deficiency anemia of children between 12 and 18 months effect their intellectual and psychomotor development, generally in women it increases the risk of stillbirth, premature delivery, and low birth-weight of children. The common treatment of
this disease is oral iron therapy as large number of iron salts are available [2].

In clinical practice, oral iron supplements commonly used are ferrous sulfate, ferrous fumarate. These typical preparations are cheap but have more gastrointestinal intolerance. To avoid that, few new iron salts have been marketed such as ferrous biglucinate and iron polymaltose complex with low gastrointestinal irritation, as the iron stores are increased the hemoglobin level also increases faster than those traditional iron salts.

While ascorbic acid is water soluble vitamin. It is said to be one of the most important ones due to its antioxidant and pH regulator properties, vitamin C is also added to food products as additive with various functional roles based on its redox properties and to various pharmaceuticals as stabilizer, modifier, and enhancer [3,4].

The new ferrous biglucinate effervescent formulation is a combination of iron and ascorbic acid where iron is in complex form with glycine (an amino acid) [5]. The valence of the ferrous ion is neutralized by 1:2 metal to ligand reaction so it restricts unwanted reactions and protects the gastrointestinal surface from irritation. Ascorbic acid is added as a supplement with iron that helps to improve the bioavailability of iron. This formulation has less gastric irritation, more bioavailability and palatable to children when compared to other commercial products.

Glycine amino acid have protective capacity with a ligand to metal ratio 2:1, each chelated Fe atom have sufficiently strong chelating bonds to resist cleavage through reactive natural foodstuff or digestion. The formed chelates protect the mineral atoms to be absorbed and used nutritionally. Supplemental levels of orally administered iron would provide more protection of gastrointestinal surface from irritation and prevent iron damage to the body when given in the form of bioavailable chelate.

Previously, Lee and Stumm has mentioned about the determination of ferrous iron in the presence of ferric iron with bathophenanthroline (BPA) and the concentration used was 0.01 M with pH 4 and hydroxylamine as reducing agent at 533 nm [6]. Syed Salman Lateef has developed an reversed phase-high-performance liquid chromatography (RP-HPLC) method for analysis of ascorbic acid, citric acid, and benzoic acid using 20 mM KH₃PO₄ of pH 2.5 with ortho phosphoric acid (OPA), methanol:acetonitrile (MEOH:ACN) (60:40) as mobile phase and column of agilent poroshell 120 EC-C18, 4.6 mm × 100 mm, 2.7 μm at 245 nm [7] and Verschoor and Molothas discussed about the evaluation of low levels of ferrous and total reactive iron using three common colorimetric methods BPA, triazine, and ferrozone in fresh waters [8]. Stumm and Lee has developed a study that determines the reaction between ferrous iron and oxygen [9] and Lazić, et al., has mentioned about the stability of 1,10 phenanthroline with hydroxylamine hydrochloride as reducing agent [10]. Adhikamsetty et al., has mentioned about kinetics of ferroxin complex with respect to Fe²⁺ by using the stopped-flow technique [11].

Indian Pharmacopoeia 2014 mentions redox titrations for ferrous fumarate, ferrous gluconate, ferrous sulfate API and iodimetric titration for ascorbic acid API and injection [12]. USP-NF 2015 mentions redox titrations for ferric ammonium citrate oral solution, ferrous fumarate, ferrous gluconate, ferrous sub sulfate, ferrous sulfate API and potentiometric redox titration for iron and also mentions iodimetry for ascorbic acid API and tablets [13].

FAO specification 2003 for ferrous biglucinate processed with citric acid has mentioned assay method by redox titration using 0.1 N cerric sulfate as titrant and orthophenanthroline as indicator [14].

No analytical method was reported so far for the analysis of iron and ascorbic acid in effervescent tablets. Hence, an attempt was made to develop and validate the colorimetric and RP-HPLC method for ferrous biglucinate effervescent tablets.

**MATERIALS AND METHODS**

**Materials and Reagents**

Ascorbic acid (sigma Aldrich), ferrous biglucinate (FBG31511007), AR grade OPA, hydrochloric acid (HCl), sulfuric acid and HPLC grade methanol, acetonitrile were purchased from Rankem. AR grade 1,10 phenanthroline, sodium acetate anhydrous were purchased from SD fine and potassium dihydrogen phosphate from Merck and an in-house Milli Q-water was used.

**Methodology**

**Colorimetric method for estimation of total iron**

The optimized conditions for the colorimetric estimation of total iron were as follows. The wave length used was 510 nm with diluent-1 as 0.5 M sulfuric acid and 1.2 M sodium acetate 5 mL. The diluent-2 was Milli Q-water. 10 mL of 1,10 phenanthroline (1 mg/mL) used as complexing agent.

Standard solution was prepared by weighing and transferring about 26.12 mg (equivalent to 5 mg of ferrous) and 50 mg of ascorbic acid to 200 mL dried volumetric flask dissolved and made up to the volume with diluent.

Placebo was prepared by weighing and transferring about 2000 mg of placebo into 200 mL dried volumetric flask, 30 mL of diluent was added and sonicated for 10 min and made up to the volume with diluent.

Sample solution was prepared by weighing and crushing 5 effervescent tablets and the tablet powder equivalent to about 2.5 mg of ferrous was weighed and transferred into 100 mL dried volumetric flask, 20 mL of diluent was added and allowed to wait till effervescence ceases. Sonicated for 10 min and made up to the volume with diluent.

In the colorimetric method development, a volume of 5 mL from the stock solution of (standard/sample/placebo) was pipette out and transferred to 50 mL volumetric flask, 5 mL of 1.2 M sodium acetate and 10 mL of 1,10 phenanthroline was added and allowed to stand for 15 min. Later it was made up to the volume with Milli Q-water.

**Method validation**

Analytical method validation was performed as per ICH Q2 (R1) [15] using Shimadzu UV 1800 at PDC, Steer Life, Bengaluru.
Specificity (interference from ferrous bisglycinate)
In this study, diluent, placebo, standard, and sample solution was analyzed in the ultraviolet range.

Linearity
Linearity was demonstrated using five determinations in the range from 25% to 150% of sample concentration. A series of ferrous bisglycinate standard preparations were prepared over a range of 0.625-3.75 μg/mL.

Accuracy
The accuracy of this method was performed by recovery study. A known amount of standard drug (analyte) was spiked in triplicate to the pre-analyzed samples, resulting mixtures were then assayed and percentage recovery of the drug was calculated. Accuracy was performed at three known concentration levels of 50%, 100%, and 150% standard concentration. All the analyses were repeated and the % recovery was calculated at each level. The mean percentage recovery values with their SDs for the proposed method were calculated.

Preparation of stock solution for accuracy
Ferrous bisglycinate standard about 130.58 mg equivalent to 25 mg of iron was weighed and taken into 100 mL dried volumetric flask and made up to the volume with diluent.

Acceptance criteria: The mean content of total iron should be 97-102%.

Precision
Method precision was determined from six independent determinations of sample solution.

Sample preparation is same as accuracy.

Acceptance criteria: The % relative standard deviation (RSD) of the six sample preparations should be ≤2%.

Analytical method development for ascorbic acid using HPLC
Agilent 1260 infinity HPLC instrument was used:

Preparation of solutions
0.3 M HCl with 5% OPA was used as a diluent to mitigate the stability problem of the sample.

Preparation of mobile Phase A (20 mM) potassium dihydrogenphosphate
About 2.72 g of potassium dihydrogenphosphate was weighed, dissolved in 1000 mL water and pH was adjusted to 2.5 ± 0 with dilute OPA and degassed.

Preparation of diluent (0.3 M HCl with 5% OPA)
About 25.5 mL of concentrated HCl was transferred to 1000 mL volumetric flask and 500 mL of water was added and mixed well to this 50 mL of OPA was added and made up to the mark with Milli-Q WATER (Table 1).

Method validation
Preparation of standard solution of ascorbic acid
About 20 mg was weighed and transferred to 100 mL dried volumetric flask dissolved and made up to the volume with diluents. 5 mL of solution was pipette out and transferred into 25 mL of volumetric flask and made up to the mark with diluent (40 μg/mL).

Preparation of placebo solution
About 2500 mg of placebo was weighed and taken into 50 mL dried volumetric flask and 30 mL of diluent was added. The solution was sonicated for 5 min and made up to the mark with diluent. The solution was filtered through Whatmann filter paper #42. From the filtrate 5 mL was pipette out and transferred into 25 mL volumetric flask and diluents were made up to 25 mL.

Preparation of sample solution
5 Tablets (equivalent to about 200 mg of vitamin C) were weighed and taken into 250 mL of dried volumetric flask and 50 mL of diluent was added and allowed to wait till effervescence ceases. The sonicated solution (for 5 min) was used to make up to the mark. After that, the solution was filtered through Whatmann filter paper #42, 5 mL of filtrate was pipette out and taken into 100 mL of volumetric flask and made up to the mark with diluent.

System suitability
Acceptance criteria:
A. The % RSD of area of Ascorbic acid peak in standard solution NMT 2.0
B. In standard solution the tailing factor should NLT 0.9 and NMT 1.5 of ascorbic acid peak.
C. Theoretical plates of Ascorbic acid peak should NLT 2000.

Specificity
In this study, diluent, placebo, standard and sample solution shall be analyzed with photo diode array detector. Moreover, interference from diluent and placebo to ascorbic acid elution in standard and sample solution shall be evaluated.

Acceptance criteria: Interference from blank and placebo peak with the main peak should NMT 0.1%.

Linearity
Linearity was performed by using minimum of six determinations in the range from 10% to 150% of sample concentration. A series of Ascorbic acid standard preparations were prepared over a range of 4 μg/mL to 60 μg/mL.

Acceptance criteria: The correlation coefficient should be NLT 0.995 ($r^2 \geq 0.990$) and the graph should be linear.

Accuracy (% recovery)
The accuracy is performed at the 25%, 50%, 75%, 100%, and 150% levels in triplicate along with placebo.

<table>
<thead>
<tr>
<th>Table 1: Optimized chromatographic conditions</th>
</tr>
</thead>
<tbody>
<tr>
<td>Column</td>
</tr>
<tr>
<td>Flow</td>
</tr>
<tr>
<td>Column temperature</td>
</tr>
<tr>
<td>Sample volume</td>
</tr>
<tr>
<td>Wavelength</td>
</tr>
<tr>
<td>Run time</td>
</tr>
<tr>
<td>Mobile phase A</td>
</tr>
<tr>
<td>Mobile phase B</td>
</tr>
<tr>
<td>Mobile phase composition</td>
</tr>
</tbody>
</table>

http://www.tjps.pharm.chula.ac.th
Preparations were filtered with Whatmann #42 and further pipette out 5 mL and made up to 25 mL with diluent.

Acceptance criteria: The mean content of Ascorbic acid must be 97-102%.

Precision
It was performed from six independent determinations of sample solution.

Preparation of sample
About 100 mg of Ascorbic acid and 2500 mg of placebo was weighed and transferred to 250 mL volumetric flask 0.30 mL of diluent was added and for 5 min the solution was subjected to sonication, finally made up to the mark with diluent.

Acceptance criteria: The %RSD for six sample preparations should be ≤2.0%.

Solution stability
The stability of standard and sample solution is evaluated at varied time points i.e., at 0 h, 2 h and 5 h. If standard and sample solution data fails at room temperature it is checked at 2-8°C.

Acceptance criteria: Recovery% should be 98-102%.

RESULTS AND DISCUSSION

Validation Summary for Ferrous Bisglycinate

Specificity results
Table 2 shows specificity results of ferrous iron.

Linearity
Table 3 shows linearity results of ferrous iron. Figure 1 shows linearity of ferrous iron complex.

Accuracy results
Table 4 shows accuracy results of ferrous iron.

Precision results
The developed analytical method was validated as per ICH guidelines Q2(R1) for specificity, linearity, accuracy, precision and the results were found to be within acceptable limits (Table 5).

Validation of Ascorbic Acid

Specificity
The sample was prepared with concentration 40 μg/mL and the area was found to be 2545.754 and the recovery with respect to standard was found to be 98.2% (Figures 2-5).

Linearity
Table 6 shows linearity results of ascorbic acid. Figure 6 shows linearity of ascorbic acid.

Accuracy results
Table 7 shows accuracy results of ascorbic acid.

Precision results
Table 8 shows precision results of ascorbic acid.
Solution stability

Weight of the standard: 20.43 mg
Weight of the sample: 50.43 mg

The developed analytical method was validated as per ICH guidelines Q2 (R1) for specificity, linearity, accuracy, precision and the results were found to be within acceptable limits. The solution stability study reveals that the solution has to be freshly prepared before use whereas both standard and sample are stable up to 5 h at 5°C (i.e., in sample port) (Table 9).

CONCLUSION

The need for the development of an analytical method was to estimate total iron and ascorbic acid in ferrous bisglycinate effervescent tablets. The developed method was validated as per ICH guidelines Q2 (R1).

By the solution stability study, it became clear that, the sample solution has to be freshly prepared before analyses.

A colorimetric method was developed for estimation of total iron with 1,10 phenanthroline at 510 nm. The developed method was validated by evaluating specificity, linearity, accuracy, and precision.
Table 6: Linearity results of ascorbic acid

<table>
<thead>
<tr>
<th>Concentration of ascorbic acid μg/mL</th>
<th>Area</th>
</tr>
</thead>
<tbody>
<tr>
<td>4</td>
<td>248.651</td>
</tr>
<tr>
<td>10</td>
<td>621.171</td>
</tr>
<tr>
<td>20</td>
<td>1244.156</td>
</tr>
<tr>
<td>30</td>
<td>1867.013</td>
</tr>
<tr>
<td>40</td>
<td>2506.870</td>
</tr>
<tr>
<td>50</td>
<td>3084.737</td>
</tr>
<tr>
<td>60</td>
<td>3732.381</td>
</tr>
</tbody>
</table>

Table 7: Accuracy results of ascorbic acid

<table>
<thead>
<tr>
<th>Sample ID</th>
<th>Wt. taken (mg)</th>
<th>Concentration μg/mL</th>
<th>Area</th>
<th>Recovery (%)</th>
<th>Average</th>
<th>SD (±)</th>
<th>(%) RSD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Acc_25%_1</td>
<td>12.88</td>
<td>10.2</td>
<td>629.749</td>
<td>99.5</td>
<td>100.8</td>
<td>1.40</td>
<td>1.39</td>
</tr>
<tr>
<td>Acc_25%_2</td>
<td>12.92</td>
<td>10.3</td>
<td>639.659</td>
<td>100.7</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Acc_25%_3</td>
<td>12.94</td>
<td>10.3</td>
<td>650.488</td>
<td>102.3</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Acc_50%_1</td>
<td>25.70</td>
<td>20.4</td>
<td>1271.143</td>
<td>100.6</td>
<td>101.7</td>
<td>0.88</td>
<td>0.87</td>
</tr>
<tr>
<td>Acc_50%_2</td>
<td>25.73</td>
<td>20.4</td>
<td>1291.894</td>
<td>102.2</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Acc_50%_3</td>
<td>25.45</td>
<td>20.2</td>
<td>1277.929</td>
<td>102.2</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Acc_75%_1</td>
<td>37.90</td>
<td>30.1</td>
<td>1870.350</td>
<td>100.4</td>
<td>100.4</td>
<td>0.17</td>
<td>0.17</td>
</tr>
<tr>
<td>Acc_75%_2</td>
<td>37.90</td>
<td>30.1</td>
<td>1873.950</td>
<td>100.6</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Acc_75%_3</td>
<td>37.66</td>
<td>29.9</td>
<td>1855.766</td>
<td>100.3</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Acc_100%_1</td>
<td>50.43</td>
<td>40.0</td>
<td>2502.929</td>
<td>101.0</td>
<td>101.7</td>
<td>0.58</td>
<td>0.57</td>
</tr>
<tr>
<td>Acc_100%_2</td>
<td>50.76</td>
<td>40.3</td>
<td>2545.754</td>
<td>102.1</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Acc_100%_3</td>
<td>50.48</td>
<td>40.1</td>
<td>2528.394</td>
<td>101.9</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Acc_150%_1</td>
<td>75.45</td>
<td>59.9</td>
<td>3743.247</td>
<td>101.0</td>
<td>101.5</td>
<td>0.69</td>
<td>0.68</td>
</tr>
<tr>
<td>Acc_150%_2</td>
<td>75.39</td>
<td>59.9</td>
<td>3754.475</td>
<td>101.3</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Acc_150%_3</td>
<td>75.81</td>
<td>60.2</td>
<td>3811.300</td>
<td>102.3</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

RSD: Relative standard deviation

Table 8: Precision results of ascorbic acid

<table>
<thead>
<tr>
<th>Sample ID</th>
<th>Wt. taken (mg)</th>
<th>Concentration μg/mL</th>
<th>Area</th>
<th>Recovery (%)</th>
<th>Average</th>
<th>SD (±)</th>
<th>(%) RSD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sample_1</td>
<td>50.43</td>
<td>40.0</td>
<td>2502.929</td>
<td>100.0</td>
<td>100.0</td>
<td>0.81</td>
<td>0.8</td>
</tr>
<tr>
<td>Sample_2</td>
<td>50.76</td>
<td>40.3</td>
<td>2545.754</td>
<td>101.1</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sample_3</td>
<td>50.48</td>
<td>40.1</td>
<td>2528.394</td>
<td>100.9</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sample_4</td>
<td>50.30</td>
<td>39.7</td>
<td>2479.277</td>
<td>99.3</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sample_5</td>
<td>50.65</td>
<td>40.1</td>
<td>2517.802</td>
<td>100.2</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sample_6</td>
<td>50.71</td>
<td>39.5</td>
<td>2493.915</td>
<td>99.1</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

RSD: Relative standard deviation

Figure 5: Chromatogram for ascorbic acid sample
Table 9: Solution stability of ascorbic acid

<table>
<thead>
<tr>
<th>Sample ID</th>
<th>Area</th>
<th>Recovery (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Standard initial_room temp</td>
<td>2534.202</td>
<td>100.0</td>
</tr>
<tr>
<td>Sample initial_room temp</td>
<td>2502.929</td>
<td>101.1</td>
</tr>
<tr>
<td>After 2 h_std._room temp</td>
<td>2096.253</td>
<td>82.7</td>
</tr>
<tr>
<td>After 2 h_sample_room temp</td>
<td>2058.479</td>
<td>82.3</td>
</tr>
<tr>
<td>After 5 h_std._room temp</td>
<td>2065.376</td>
<td>81.5</td>
</tr>
<tr>
<td>After 5 h_sample_room temp</td>
<td>2291.377</td>
<td>91.6</td>
</tr>
<tr>
<td>After 2 h_std._5°C</td>
<td>2495.073</td>
<td>98.5</td>
</tr>
<tr>
<td>After 2 h_sample_5°C</td>
<td>2487.035</td>
<td>99.4</td>
</tr>
<tr>
<td>After 5 h_std._5°C</td>
<td>2502.862</td>
<td>98.8</td>
</tr>
<tr>
<td>After 5 h_sample_5°C</td>
<td>2472.401</td>
<td>98.9</td>
</tr>
</tbody>
</table>

Figure 6: Linearity of ascorbic acid

Chromatographic separation was developed on a Inertsil C_{18} (250 mm × 4.6 mm, 3 μm) column at 245 nm by using an isocratic program for 10 min. The mobile phase includes 20 mM potassium dihydrogen phosphate (pH 2.5). The developed method was validated for specificity, linearity, accuracy, and precision.

The results demonstrate that the proposed method is simple and more economical in comparison with other reported methods discussed in literatures and it has all the potential for routine quality control analysis.

ACKNOWLEDGMENTS

The authors are gratefully acknowledge Steer Life India Ltd., Bengaluru, Karnataka, India and Department of Pharmaceutical Quality Assurance, Manipal College of Pharmaceutical Sciences, Manipal University Manipal, Karnataka, India for facilities to carry out the research work.

REFERENCES