Genetic Heterogeneity of Kidney Stone Disease in Northeastern Thai Patients

Choochai Nettuwakul¹, Oranud Praditsap², Nunghathai Sawasdee¹, Thanakorn Pungsrinont¹, Suchai Sritippayawan³, Nawara Faiza Ahsan^{1,4}, Pa-thai Yenchitsomanus¹, Nanyawan Rungroj^{2,*}

¹Division of Molecular Medicine, Research Department, Faculty of Medicine Siriraj Hospital, Mahidol University, Bangkok,10700, Thailand

²Siriraj Genomics, Office of the Dean, Faculty of Medicine Siriraj Hospital, Mahidol University, Bangkok, 10700, Thailand

³Division of Nephrology, Department of Medicine, Faculty of Medicine Siriraj Hospital, Mahidol University, Bangkok, 10700, Thailand

⁴Immunology Graduate Program and Department of Immunology, Faculty of Medicine Siriraj Hospital, Mahidol University, Bangkok 10700, Thailand

*Corresponding author: nrungroj@yahoo.com

ABSTRACT

Kidney stone disease (KSD) is a health problem worldwide, with a reportedly increasing prevalence and incidence across the world. In Thailand, KSD is common in the north (N) and northeastern (NE) parts of the country, with the NE region accounting for more than 40% of total patients. Calcium stones comprise 80% of all kidney stones, making them the most prevalent globally, as well as in Thailand. Generally, KSD is often associated with metabolic abnormalities of the urinary solute concentration or decreased urinary solubility, including hyperoxaluria, hypercalciuria, hypocitraturia, hyperuricosuria, cystinuria, low urinary volume, and defects in urinary acidification. The etiology of KSD is poorly understood, but it is known that both genetic and environmental factors are involved and the disease is heterogeneous, ranging from monogenic defects to complex interactions between genetic and environmental factors. Reported studies of KSD using genetic and genomic approaches have revealed that ion transporters and channels, the calcium-sensing receptor signaling pathway, and the metabolic pathways of vitamin D, oxalate, cysteine, purines, and uric acid play important roles in causing KSD. In Thailand, studies of KSD employing different genetic and genomic approaches i.e. candidate gene analysis, genome-wide association studies (GWAS), linkage analysis, and next-generation sequencing, have identified genetic alterations in F2, PAQR6, SLC13A2, ITLN1, SCN10A, and PBK genes. These have provided insights into both the common and rare genetic variants and

genetic heterogeneity of KSD in Thai patients. The identification of genetic defects and molecular pathways causing KSD in the Thai cohort contributes to the increase of our understanding of the pathogenesis of KSD.

Keywords: kidney stone disease; single nucleotide polymorphism; exome sequencing; genetic heterogeneity; Northeast Thailand

INTRODUCTION

Kidney stone disease (KSD) is a condition in which insoluble material is formed in the kidney from the crystallization of supersaturated substances in urine, which may adhere and deposit at the tubular epithelial cells in the kidney or pass through the urinary tract. Calcium stones are the most common stone type, comprising 80% of all stones, globally. However, other types of stones including uric acid, struvite, cystine, and xanthine have also been found (Coe *et al.*, 2005; Lieske *et al.*, 2014). Similar to the global trend, the main stone types found in Thai patients are calcium stones (Aegukkatajit *et al.*, 1994; Yanagawa *et al.*, 1997).

KSD is a major health problem with increasing prevalence and incidence across the world (Romero *et al.*, 2010). The prevalence of kidney stones is 5%-15% in the world populations, with a recurrence rate of 50% within 5–10 years, and 75% in 20 years (Johnson *et al.*, 1979; Trinchieri *et al.*, 1999). The high recurrence rates lead to higher costs of medical treatment, hospitalization, healthcare

expenses, social costs, and indirect costs from time lost from work, thus creating an economic burden (Saigal *et al.*, 2005). In the United States, the male to female ratio of KSD prevalence was reported to be 10.6%:7.1% (Scales *et al.*, 2012). In Thailand, kidney stones are particularly common in the north (N) and northeastern (NE) parts of the country, where their prevalence is 6.6% (Sritippayawan *et al.*, 2009). The incidence rate in Thai population is higher than 85 per 100,000. Of these cases, more than 40% are from the NE region (National Statistical Office, 2021).

In the patients, KSD can be both asymptomatic and symptomatic. Usually, the first symptom of a kidney stone is extreme pain, which may begin suddenly when the stone attempts to pass through the urinary tract, blocking the flow of urine. Other symptoms associated with kidney stones are flank pain, hematuria, frequent painful urination, nausea, vomiting, and fever (Coe et al., 2005). The etiology of kidney stones is unclear but both genetic and environmental factors are involved. Thus, KSD is heterogeneous and its etiology ranges from monogenic defects to complex interactions between genetic and environmental factors (Gambaro et al., 2004). KSD is considered to be a systemic disorder associated with other non-communicable diseases, such as chronic kidney disease, renal failure, coronary artery disease, hypertension, type 2 diabetes mellitus, and metabolic syndrome (Madore et al., 1998; Rule et al., 2009; Rule et al., 2010). It is often associated with metabolic abnormalities of the urinary solute concentration or decreased urinary solubility, including hypercalciuria, hyperoxaluria, and hyperuricosuria. Notably, the clinical characteristics of the disease in NE Thai patients were not similar to those previously reported in other ethnic groups, whereby hypocitraturia and potassium deficiency were observed predominantly in these patients (Sriboonlue et al., 1991). Therefore, it was hypothesized that the mechanisms of stone formation in this population may differ from those previously reported in other ethnic groups.

The mechanisms involving in stone formation are the interaction of crystals in supersaturated urine with some intra-renal structure, crystal growth, and crystal aggregation, consequently leading to stone formation (Aggarwal *et al.*, 2013). The risk factors for stone formation include genetic and environmental factors. The most common types of stones are calcium oxalate stones. The conditions that increase urinary calcium and oxalate excretion enhance calcium oxalate supersaturation will therefore increase the risk of KSD. Additionally, factors such as calcium and oxalate intake, gastrointestinal tract (GI) absorption, bone turnover, and renal reabsorption influence the calcium and oxalate levels in both serum and urine. These processes are controlled by the parathyroid hormone, 1,25 dihydroxy vitamin D, and calciumsensing receptor (CaSR) signaling, whose alterations can act as risk factors of KSD (Howles et al., 2019). There is evidence suggesting that vitamin D receptor polymorphism is associated with the (VDR)hypercalciuria phenotype and a higher risk of stone formation (Mossetti et al., 2004; Bid et al., 2005). Moreover, several variations in the CaSR gene cause a decrease in CaSR expression and kidney stone development (Vezzoli et al., 2011). Food intake of oxalate and its intestinal absorption as well as oxalate metabolism in the liver or kidney also contribute to the hyperoxaluria underlying and KSD. Primary hyperoxaluria is a severe metabolic disorder. characterized by a deficiency or mistargeting of specific enzyme in the glyoxalate metabolic pathway, resulting in excessive oxalate production (Danpure and Rumsby, 2004; Cellini et al., 2012; Beck et al., 2013). Patients with digestive diseases, such as inflammatory bowel disease, can develop secondary hyperoxaluria, leading to kidney stone formation (Torricelli et al., 2021). Moreover, Oxalobacter formigenes in humans have been proposed to participate in intestinal oxalate metabolism (Allison et al., 1986; Stewart et al., 2004). Other calcium oxalate stone promoters are high sodium, high urate, low urine pH, and low urine volume, while the inhibitors include organic substances such as nephrocalcin, urinary prothrombin fragment-1, and osteopontin and inorganic substances such as citrate and magnesium (Aggarwal et al., 2013). The mechanism of kidney stone formation includes calcium oxalate monohydrate (COM) crystals adhesion or endocytosis on to renal tubular epithelial cells (Bigelow et al., 1996; Lieske et al., 1999; Evan et al., 2005). The oxalate, COM, and CaP exposure to renal tubular cells increases the production of reactive oxygen species (ROS) and oxidative stress, leading to cell injury and inflammation. This accelerates crystal aggregation and growth, which ultimately leads to stone formation (Khaskhali et al., 2009; Khan, 2013).

In this review, we focus on the genetic and genomic studies of KSD in the NE Thai population, including case-control and family-based studies, to provide an update on the genes associated with KSD and the related mechanisms of stone formation. The accumulative data gathered from our studies indicate the genetic heterogeneity of KSD in the NE Thai patients.

Epidemiology and genetic studies of KSD

Many studies have demonstrated that individuals with family history of KSD have a higher risk of developing the disease than those without family history. Approximately 40% of the patients with KSD have a positive family history, in which the onset of the disease is earlier than that without the family history (Koyuncu et al., 2010). Twin studies have estimated the heritability of the risk for KSD to be 56%-57% in men, and 46% in women (Goldfarb et al., 2005; Halbritter et al., 2015); thus, suggesting the role of genetic factor involving in KSD. Furthermore, epidemiological studies have estimated the relative risk (λ_R) in family members to range from 2-4-fold up to 1 6 -fold higher than that of the general population, depending on the degree of relatedness (Griffin, 2004). In Thailand, KSD is prevalent in the N and NE regions of the country. The reported prevalence of KSD in this population is considerably varied among previous

studies (Sriboonlue et al., 1992; Nimmannit et al., 1996; Yanagawa et al., 1997), which is probably related to the method conducted in the different studies. According to the Office of the Permanent Secretary, Ministry of Public Health there are more than 50,000 urolithiasis patients /year in Thailand and the incidence rate in the population is more than 85 per 100,000. Of these cases, more than 40% are from the NE region (Figure 1A and 1B) (National Statistical Office, 2021). The reason for such higher susceptibility in the NE region of Thailand is still unknown. However, there are evidence suggesting low potassium and low citrate status in the rural dwellers of NE, due to low daily dietary intake and loss through sweat, as risk factors (Sriboonlue et al., 1998; Tosukhowong et al., 2002). In addition, genetic factors play some roles in the pathogenesis of KSD in the region. A study conducted in Khon Kaen province showed the relative risk ($\lambda_{\rm R}$) among family members to be 3.18 (Sritippayawan et al., 2009).

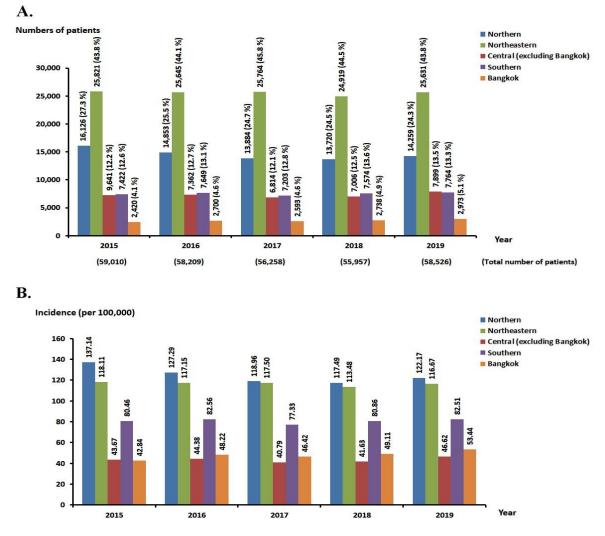


Figure 1 Number of patients (A) and incidence of kidney stone disease (KSD) in Thailand (B). The data in four regions were collected between 2015–2019.

The previous studies suggested that the probable mode of inheritance of KSD is autosomal dominance with variable or incomplete penetrance (Coe et al., 1979; Nicolaidou et al., 1996; Reed et al., 1999; Wolf et al., 2005) or the mixture of codominant and polygenic transmission (Loredo-Osti et al., 2005). Regarding to the mode of inheritance, KSD could be inherited as a rare monogenic disorder or complex polygenic transmission. This implies that many genes may contribute to the disease causation. The monogenic causes of KSD are verv heterogeneous. Genes responsible for hypercalciuria, hyperoxaluria, and other risk phenotypes have been reported. For example, mutations in CLDN16, CLCN5, SLC34A3, CaSR, SLC12A1, KCNJ1, ClCNKB, and ADCY10 genes cause KSD related to hypercalciuria (Sayer, 2017; Halbritter, 2021), whereas mutations in AGXT, GRHPR, and HOGA1 genes can cause KSD associated with primary hyperoxaluria (Danpure and Rumsby, 2004; Cellini et al., 2012; Beck et al., 2013). Mutations in at least 30 genes have been identified as monogenic genes of KSD and/or nephrocalcinosis. Recent reports of KSD in European and European American cohorts suggested that monogenic disorders account for 16.8%-20.8% of the cases in children and 11.4% of the cases in adults (Halbritter et al., 2015; Braun et al., 2016). Recessive monogenic mutations occur more commonly in children, especially in the infantile subgroup. In contrast, dominant monogenic mutations are found more frequently in patients with late-onset KSD (Halbritter et al., 2015; Daga et al., 2018). Interestingly, mutations in the cystinuria gene SLC7A9 were most frequently found in an American cohort, with median age of stone manifestation at 26 years (Halbritter et al., 2015). Moreover, clinical data showed possible cystinuria and some patients exhibited calcium-containing kidney stones (Cupisti et al., 2012; Halbritter et al., 2015). The contributions of polygenic influences from multiple loci have been investigated by candidate gene and genome-wide association studies in various populations, which have indicated that multiple genes and molecular pathways contribute to the risk of kidney stone formation. Polymorphisms in multiple loci have been reported to be associated with KSD: VDR, ALPL, SLC34A1, SLC34A4, CLDN14, TRPV5, CYP24A1, and CaSR genes encoding proteins for the renal handling of phosphate and calcium; F2, MGP, OPN, HSPG2, PLAU, and UMOD genes encoding proteins for preventing calcium salt precipitation or inhibiting stone formation; AOP1 gene encoding protein for a

water-specific channel in the kidney proximal tubule (Taguchi *et al.*, 2017; Halbritter, 2021).

Genetic and genomic approaches for the identification of genes associated with KSD

To date, several approaches have been applied for the identification of disease-causing or diseaseassociated genes in Mendelian disorders or complex diseases. Each approach provides both advantages and limitations (Table 1), which researchers can appropriately select for their work depending on their study design, sample collection, disease knowledge, budget, and application. To identify genes responsible for KSD, genetic approaches based on genetic association study, linkage study, and exome sequencing are employed. In contrast to the traditional association study and linkage study approach, genome-wide association study (GWAS) and genome-wide linkage analysis were developed as rapid, high-throughput, and cost-effective alternative approaches. In addition, these are hypothesis-free approaches to identify the disease locus in a single run. By using single nucleotide polymorphisms (SNPs) as genetic markers, the functional SNPs can be further defined. Usually, the identified disease locus is too large for conventional Sanger sequencing, making it unable to reduce the number of candidate genes sufficiently (Botstein and Risch, 2003). This problem can now be solved by the development of recent exome sequencing technologies, i.e., the next-generation sequencing, to sequence the DNA region of interest. Moreover, exome sequencing can be used to identify all kinds of genetic variations at base-pair resolution throughout the coding sequences of the human genome (Gilissen et al., 2011).

Association study

Genetic association study is a powerful method used to identify the susceptible genes or genome regions that contribute to common and complex diseases. This approach is based on the principle of linkage disequilibrium (LD), in which common variations are selected for genotyping (Table 1). Genetic association studies generally aim to identify the genetic variation that occurs more often in individuals with the disease (case group) than in individuals without the disease (control group). To investigate the correlation between disease status and genetic variation, a higher frequency of the genetic markers in the case group can be interpreted as an increased risk of a disease. Genetic association study findings have been used to explain a small effect of the causes in complex non-Mendelian diseases

(Farber and Lusis, 2009) or the sum of the phenotypic effect of multiple genes. The availability of SNP microarray allows researchers to perform an association study at a genome scale and to exponentially increases the ability to identify phenotype–genotype associations (MacArthur *et al.*, 2017). Using this approach, a number of genetic variations have been reported to be associated with KSD. Here, we summarize genetic etiology of KSD in some Asian populations, identified by association studies (Table 2).

Linkage analysis

Linkage analysis is another powerful method for identifying potential pathogenic variants, and is applicable to both monogenic diseases (parametric linkage) and complex diseases (modelfree or non-parametric linkage) (Dawn Teare and

Barrett, 2005). The principle of this approach is based on the law of segregation, which states that allele pairs segregate randomly from each other during gamete formation (Table 1). Using common variants as genetic markers, linkage analysis can be used to trace the gene that co-segregates with a disease trait without actually knowing mutation. Known mutations can also be selected for genotyping in family members by linkage analysis. Identification of potential genetic contributions with small effects can be accomplished, owing to recent genotyping technology and statistical strategies (Bodmer and Bonilla, 2008). Mutations that segregate with KSD have been identified by linkage analysis, including mutations in the SCN10A (Nettuwakul et al., 2018), SLC34A3 (Tencza et al., 2009), and OCRL1 (Hoopes et al., 2005) genes.

Table 1 The features and characteristics of association study, linkage analysis, and next generation sequencing for the identification of genes associated with the disease.

Features	Association Study	Linkage Analysis	Next Generation Sequencing
Biological Basis	Linkage disequilibrium (LD) and recombination events in past generations of a population	Segregation and recombination of genetic markers in pedigree data	Massively parallel DNA sequencing to determine variants
Hypothesis for disease causing	Hypothesis based for candidate gene study and hypothesis free for genome- wide study	Same as association study	Both hypothesis based and hypothesis free approach possible
Subjects	Population studies in affected subjects (assumed unrelated cases and controls)	Pedigree analysis among family members	Related or unrelated individuals
Benefits	 Large scale genotyping to detect common variants in a population Do not need familial or relationship information Can identify multiple genes each with minimal effects 	 Good for monogenic Mendelian diseases Can identify rare alleles that are present in small numbers of families Can be applied to multifactorial disease locus mapping 	 Better coverage and resolution to identify direct causal genes Detection of rare causal variants Applicable to both pedigree and population studies
Limitations	 Requires large sample to establish significant association in genes with small effect Prone to false positives Population stratification influences results of analysis Low power in detecting allelic heterogeneity 	 Cannot detect rare variants with low penetrance alleles May fail to replicate/ reproduce results in different families Usually have poor genetic resolution in the centimorgan range 	 High rate of discovery and need validation Storage and processing of large data sets Lack of proper standards across different NGS platform.

Country	Gene	Variation of interest	Sample size	Odds	P-Value	Reference	
			(case/control)	Ratio			
China	IL-18	rs549908	272/104	3.10	< 0.0010	Lai et al., 2010	
	VDR	rs7975232	464/450	2.04	0.0060	Wang et al., 2012	
	Klotho	rs3752472	426/282	1.51	0.0430	Xu et al., 2013	
	SLC26A6	rs184187143	225/201	6.10	0.0070	Lu et al., 2016	
	CaSR	rs1042636	615/315	1.45	0.0210	Ding et al., 2017	
	ALPL	rs1256328	331/553	1.52	0.0009	Li et al., 2018b	
	CaSR	rs7652589	624/470	1.68	0.0030	Li <i>et al.</i> , 2018a	
Japan	MGP	rs4236	122/125	0.55	0.0470	Gao et al., 2007b	
	OPN	SNP10	126/214	1.64	0.0260		
		(Novel SNP in promoter)				- Gao <i>et al.</i> , 2007a	
	OPN	SNP11	126/214	1.76	0.0113	- Guo er un, 2007u	
	(Novel SNP in promoter)						
	RGS14	rs12654812	601/201	1.43	0.0031		
	DGKH	rs7981733	601/201	1.41	0.0050	Yasui <i>et al.</i> , 2013	
	FAM188	rs12669187	601/201	1.57	0.0064	_	
Thailand	F2	rs5896	132/126	0.49	0.0030	Rungroj et al., 2012	
	PAQR6	rs759330	216/216	2.02	0.0001	Rungroj et al., 2014	
	SLC13A2	rs11567842	145/115	8.34	< 0.001	Udomsilp et al., 2018	
	ITLN1	rs2274907	216/216	1.54	0.0021	Pungsrinont et al., 2021	

Table 2 Genetic etiology of KSD in some Asian populations identified by association studies.

Next-generation sequencing (NGS)

Next-generation sequencing (NGS) is an evolution of sequencing technology. It provides massively parallel, high-throughput DNA or RNA sequencing. In contrast to the first-generation sequencing, NGS yields massive sequence data and minimizes the need for multi-steps in Sanger sequencing, such as a sample preparation step, DNA template, and sequence read length (Table 1). Most of the next-generation sequencing platforms rely on sequencing-by-synthesis technique, and conduct sequencing and detection simultaneously. Raw data received from NGS are shorter than those received from Sanger sequencing as they are determined by a function of the signal-to-noise ratio. NGS can be used for sequencing of whole genomes (genome sequencing) and targeted regions of interest (targeted sequencing or gene panel sequencing), including all coding gene sequences (exome sequencing). Using whole exome sequencing and gene panel sequencing, a number of novel and known mutations in AGXT, ATP6V1B1, CLDN16, CLDN19, GRHPR, SLC3A1, SLC12A1, SLC9A3R1,SLC34A1, SCN10A, and PBK genes have been identified as monogenic causes of KSD (Halbritter et al., 2015; Braun et al., 2016; Daga et al., 2018; Nettuwakul et al., 2020).

Genetic and genomic studies of KSD in Thailand

As mentioned earlier, almost half of the patients with KSD in Thailand can be found in the NE region of the country; therefore, many studies have recruited case subjects for clinical and genetic studies from this region. For genetic and genomic studies, several approaches have been applied to identify the genes responsible for KSD in the NE Thai population, especially candidate gene and genome-wide association studies. Moreover, most recent strategies with the NGS technology have been applied to search for the causative genes in affected families using exome sequencing.

Association studies

An overview of the genetic association studies on KSD in the NE Thai populations is provided in Table 3. The first study on the genetics of urinary stoneinhibitors in the NE Thai population was carried out by our group (Rungroj *et al.*, 2011). Urinary stone-inhibitor proteins defects including F2 (prothrombin) were proposed to be involved in kidney stone formation. The case subjects (164 cases) were patients (aged 20–80 years old) diagnosed with KSD in Khon Kaen Regional Hospital during 2004–2006. The healthy control subjects (216 controls) were people without a history of KSD and were recruited from the same area as the cases. We genotyped 67 SNPs in 8 candidate genes by dHPLC and found that 8 of 10 SNPs distributed within the *F2* gene, including 2 haplotypes, were associated with KSD risk (Figure 2). The functional SNP in the *F2* gene was further determined by Sanger sequencing. Later, an association between an *F2* variant (rs5896, p.T165M) and KSD in the NE Thai female patients (132 cases vs. 126 controls) was reported (Table 3) (Rungroj *et al.*, 2012). The significant differences in the genotype and allele frequencies were maintained only in the female group (p = 0.033 and 0.003; OR = 0.49 and 0.59, respectively), suggesting a potential protective role of *F2* against stone formation.

In addition, another study investigated the association of *SLC13A2* (*sodium dicarboxylate cotransporter-1*) polymorphism with the hypocitraturic phenotype in Thai patients with KSD (Udomsilp *et al.*, 2018). Citrate is a stone inhibitor that forms complexes with calcium (Chow et al., 2004). This study showed that SNP rs11567842 in the *SLC13A2* gene is not likely to be a direct genetic risk factor for kidney stone formation. However, it was strongly associated with the hypocitraturic phenotype in KSD patients (Table 3 and Figure 2). The patients with the AA genotype had significantly lower urinary citrate levels than those with the GG genotype. The AA genotype was susceptible to

the hypocitraturic trait, whereas GG was a protective genotype for hypocitraturia. Patients with AA had a higher expression level of *SLC13A2* mRNA in their affected kidneys than those with GG.

For whole genome SNP genotyping, our group analyzed a set of 104 reported candidate genes involved in KSD, and found that rs759330 is a potential candidate associated with KSD in the NE Thai patients (Rungroj et al., 2014). The SNP rs759330 is located 144 bp downstream of BGLAP, at a predicted microRNA binding site at 3'UTR of the PAQR6 gene. The SNP rs759330 was genotyped in 216 patients and 216 control subjects and significant differences were found in its genotype and allele frequencies (p = 0.0007 and 0.0001, OR = 2.02 and 2.02, respectively) (Table 3 and Figure 2). In the study by Pungsrinont et al., whole genome SNP genotyping was performed. SNPs were analyzed as on the concept of linkage clusters based disequilibrium (LD) and haplotypes, to identify the candidate gene associated with KSD in the NE Thai patients (Pungsrinont et al., 2021). Moreover, SNP rs2274907 in the ITLN1 gene was significantly different between the case and control subjects, in both the genotype frequencies and allele frequencies (p =0.001 and 0.0021; OR = 2.44 and 1.54, respectively) (Table 3 and Figure 2).

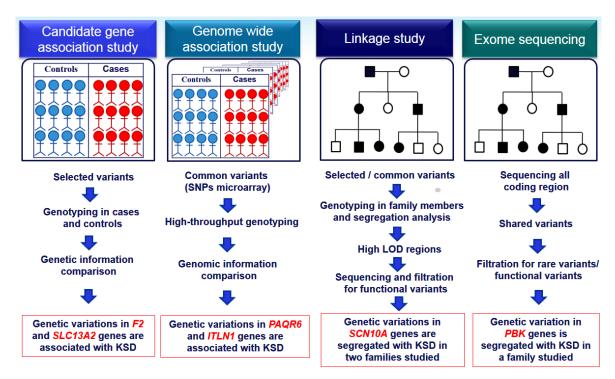


Figure 2 Genetic and genomic approaches for identification of genes associated with kidney stone disease in the northeastern (NE) Thai patients.

Family-based study

Genome-wide linkage analysis and/or exome sequencing were employed to identify disease-causing genes in KSD families by our group (Table 4). We recruited 256 patients with KSD and their family members from Sappasithiprasong Hospital in Ubon Ratchathani province. An extended family (Family 1) with a maximal estimated LOD score (ELOD) of 3.31, comprising 17 family members (7 affected and 10 unaffected), was selected for whole genome linkage analysis and exome sequencing. Two variations (c.2727C>A; p.N909K and c.5426A>G; p.K1809R) in the *SCN10A* gene were found to be segregated with KSD in Family 1 and an additional variation (c.3445G>A; p.V1149M) was identified by gene scanning in another affected family (Family 2) (Figure 2 and 3) (Nettuwakul *et al.*, 2018). More recently, we discovered a novel loss-of-function alteration (c.127G>A; p.G43R) in the *PBK* gene, which segregated in another affected family (Family 3), via exome sequencing (Nettuwakul *et al.*, 2020). This family comprised 8 affected members, including a twin pair and 20 unaffected members (Figure 2 and 3) with a maximal ELOD of 2.66.

Genes	Variations	SNP ID	Frequency (1000genome)	No. of case/ control	p-value (odd ratio)	Gene functions	Proposed KSD mechanisms	References
F2	p.T165M (c.494 C>T)	rs5896	0.2192	132/126	0.0030 (0.49)	Coagulation factor	Stone inhibitor	Rungroj et al., 2012
PAQR6	c.*395C>G	rs759330	0.2270	216/216	0.0001 (2.02)	Receptors for progesterone	Ca ²⁺ reabsorption	Rungroj et al., 2014
SLC13A2	p.1550V (c.1795A>G)	rs11567842	0.4734	145/115	<0.001 (8.34)	Sodium dicarboxylate cotransporter, citrate reabsorption	Citrate reabsorption, urinary inhibitors of stone formation	Udomsilp et al., 2018
ITLNI	V109D (c.326T>A)	rs2274907	0.4187	216/216	0.0021 (1.54)	Binding to oxalate degrading bacteria	Oxalate absorption	Pungsrinont et al., 2021

Table 3 Genetic variations identified by case/control studies.

Table 4. Genetic variations identified by family-based studies.

Genes	Variations	SNP ID	Frequency (ExAC)	Segregation In family	Function Predictions*	Gene functions	Proposed KSD mechanisms	References
SCN10A	p.N909K (c.2727C>A)	rs567269429)	0.00009	Family 1	3	Voltage-gated sodium channel	Ion imbalance	Nettuwakul et al., 2018
	p.K1809R (c.5426A>G	rs561166361)	0.000008	Family 1	6	Voltage-gated sodium channel	Ion imbalance	Nettuwakul et al., 2018
	p.V1149M (c.3445G>A)	rs560631745)	0.00007	Family 2	6	Voltage-gated sodium channel	Ion imbalance	Nettuwakul et al., 2018
РВК	p.G43R (c.127G>A)	-	-	Family 3	5	Phosphorylation of p38 MAPK	Response to ROS	Nettuwakul et al., 2020

* Prediction of the impacts of amino acid changes on the protein structures and functions by 6 web-based programs. The results show the number of programs that predict as "Pathogenic", "Damaging", "Probably damaging", "Possibly damaging", "Disease causing", or "Deleterious".

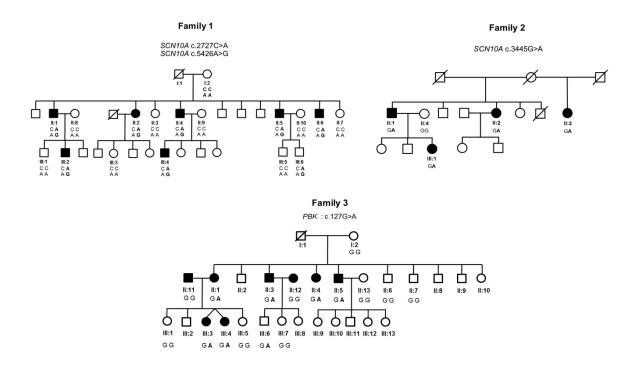


Figure 3 Segregation analyses of genetic variations in three families affected with kidney stone disease (KSD).

Functions of the genes and encoded proteins associated with KSD in Thai population

The current evidence of the potential causative genes and their mutations and polymorphisms associated with KSD in Thai population, especially in the NE Thai patients, have been collected and the results demonstrate the genetic heterogeneity of KSD in the NE Thai population. The identified genes are involved in various mechanisms of stone formation, including stone inhibition, ion imbalance, oxidative stress, and oxalate degrading bacteria as described in Figure 4 and summarized below.

F2

Prothrombin (coagulation factor II) is encoded by the F2 gene. Urinary prothrombin fragment 1 (UPTF1), the product of prothrombin, was initially described as a crystal matrix protein within calcium oxalate crystals, and is considered to be a potent inhibitor of calcium oxalate growth and aggregation in urine (Suzuki et al., 1994; Stapleton and Ryall, 1995) (Figure 4). The association study of F2 variant (rs5896 or T165M) showed that C allele was the protective allele only in the female group. The reason why rs5896 plays a role only in the female group is still unclear. However, the prevalence of KSD in the NE Thai population has been shown to be predominant in the male group, with male to female ratio of about 2:1 (Sritippayawan et al., 2009). Gender and sex hormones are known to have some influence on the prevalence of KSD. Thus, it is possible that different modifiers might differently influence males and females, or this might be the result of a complex interaction between F2 and specific sex hormones in either group. Furthermore, the UPTF1 level was high in the urine samples from female subjects carrying homozygous-CC. This could explain the protective effect of C allele in the female group with a lower risk of developing KSD (Rungroj *et al.*, 2018).

PAQR6

PAQR6 is the gene encoding progestin and adipoQ receptor family member 6, which belong to the progestin and AdipoQ receptor family (PAQR) that can be activated by progesterone or by some of its metabolites such as dihydroprogesterone and allopregnanolone. These receptors are involved in reproduction, development, immunological, and neuroendocrine responses (Fernandes et al., 2005; Dressing and Thomas, 2007; Mesiano et al., 2011; Pang et al., 2013). However, PAOR6 has never previously been described to be associated with KSD. It was hypothesized that PAQR6 may be regulated by progesterone and may simulate Ca2+ reabsorption at the distal part of the nephron (Shughrue et al., 1988; Brunette and Leclerc, 2002; Tang et al., 2005). The SNP rs759330 that is located at a predicted microRNA binding site at 3' UTR of the PAQR6 gene (Rungroj et al., 2014) might affect the level of gene expression and is correlated with Ca²⁺ reabsorption in the kidney (Figure 4).

SLC13A2

SLC13A2, encoding a sodium-coupled citrate transporter, was reported to play a role in the formation of kidney stones. Citrate inhibits calcium oxalate crystallization by forming complexes with calcium (Chow *et al.*, 2004) (Figure 4). The patients with AA genotype of SNP rs11567842 had a high expression of *SLC13A2* mRNA in their affected kidneys (Udomsilp *et al.*, 2018). A higher expression level of the *SLC13A2* gene in renal tubular cells can lead to an increased reabsorption of citrate, as well as a risk of hypocitraturia. Persistent and long-standing hypocitraturia may gradually promote the development of KSD.

ITLN1

ITLN1, encoding the intelectin-1, can specifically bind to microbial carbohydrate chains in a calcium-dependent manner, as in the case of innate immune lectins (Tsuji *et al.*, 2001; Wesener *et al.*, 2015). *ITLN1* functions which associate with KSD are still unknown. It was hypothesized that intelectin-1 may recognize bacteria in the intestinal tract. Since human intelectin-1 is highly expressed in the intestinal tract, it might be associated with a Gram-negative anaerobic bacterium like *Oxalobacter formigenes*, which plays an important role in degrading oxalate in the intestinal tract.

SCN10A

SCN10A, encoding the Na_v1.8 α subunit of the voltage-gated sodium channel, was initially reported to be preferentially expressed in peripheral sensory neurons

and heart tissue. Gain-of-function alterations of the SCN10A cause painful peripheral neuropathy (Garrison et al., 2014), whereas its loss-of-function alterations result in prolonged cardiac conduction disease and Brugada syndrome (Chambers et al., 2010; Hu et al., 2014). The Na_V1.8 α subunit protein was also found to be expressed in proximal tubules and in the collecting ducts of the nephron in the human kidney. The variant protein (p.N909K and p.K1809R in the same polypeptide chain) expressed in cultured cells was unstable, which reduces the current density, as studied by the whole-cell patch clamp technique (Nettuwakul et al., 2018). These analyses showed that loss-of-function alterations of SCN10A might reduce Na⁺ reabsorption, resulting in a high Na⁺ filtration, and decreased Ca²⁺ reabsorption (Figure 4).

PBK

The *PBK* gene, encoding the PDZ binding kinase, is a member of the mitogen-activated protein kinase (MAPKK) family, and its activated form phosphorylated p38 MAPK (Abe et al., 2000; Gaudet et al., 2000; Matsumoto et al., 2004; Ayllon and O'Connor, 2007). The p.G43R substitution results in instability of PBK and reduces the the phosphorylation of p38 MAPK (Nettuwakul et al., 2020), thereby regulating the downstream signaling pathway, including cell viability and apoptosis. This phenomenon might affect cell survival from oxidative stress, resulting in renal epithelial injury and kidney stone formation (Figure 4).

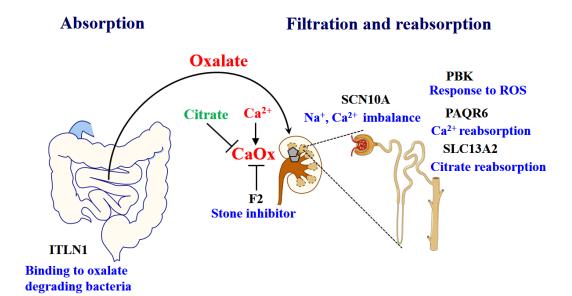


Figure 4 Genes responsible for kidney stone disease (KSD) in the NE Thai patients and their related mechanisms.

CONCLUSION

Several approaches can be applied to identify the genes responsible for KSD in Thai populations. The use of common SNPs for candidate-gene or genome-wide association approaches require further analysis to identify the causative mutations or functional SNPs. The advent of NGS technology has greatly assisted the identification of genes and mutations responsible for KSD. In this review, we presented the current evidence of the potential causative genes and mutations for KSD in the NE Thai patients. At least 4 common SNPs in F2, PAQR6, SLC13A2 and ITLN1 genes were identified by genetic association studies. Four mutations or non-synonymous alterations in SCN10A and PBK genes were identified by exome sequencing and gene scanning in the families affected with KSD. These results illustrate the genetic heterogeneity of KSD in the population in the NE region of Thailand. The identified genes are involved in various mechanisms of stone formation, such as stone inhibition, ion imbalance, oxidative stress, and oxalate degrading bacteria (Figure 4). The genes or loci identified in Thai patients with KSD provide insights into the pathogenesis of this disorder. However, additional studies are required to completely understand genetic alterations and mechanisms of KSD in the NE Thai population so that the more effective methods for prevention, treatment, and control of KSD in this population will be developed and applied.

ACKNOWLEDGEMENTS

Our research activities on the genetic and genomic studies of kidney stone disease were financially supported by Siriraj Research Fund, Faculty of Medicine Siriraj Hospital, Mahidol University (Grant No. R016034007), a Mahidol University Grant (No. R015910004), and a grant from the Kidney Foundation of Thailand. NFA was supported by Siriraj Graduate Scholarship and NR was supported by a Chalermphrakiat Grant from the Faculty of Medicine Siriraj Hospital, Mahidol University, Bangkok, Thailand.

REFERENCES

Abe Y, Matsumoto S, Kito K, Ueda N. Cloning and expression of a novel MAPKK-like protein kinase, lymphokine-activated killer T-cell-originated protein kinase, specifically expressed in the testis and activated lymphoid cells. *J Biol Chem. 2000;* 275(28): 21525-21531.

- Aegukkatajit S, Nagaphant A, Nuhung R, Sinturat R, Nugoonsawat P, Mungmai P. Epidemiological study of urinary stones based on operative theater data at regional hospitals and general hospitals of public health region-5, Thailand. J Med Assoc Thai. 1994; 77(9): 484-487.
- Aggarwal KP, Narula S, Kakkar M, Tandon C. Nephrolithiasis: molecular mechanism of renal stone formation and the critical role played by modulators. *Biomed Res Int. 2013*; 2013: 292953.
- Allison MJ, Cook HM, Milne DB, Gallagher S, Clayman RV. Oxalate degradation by gastrointestinal bacteria from humans. *J Nutr. 1986;* 116(3): 455-460.
- Ayllon V, O'Connor R. PBK/TOPK promotes tumour cell proliferation through p38 MAPK activity and regulation of the DNA damage response. *Oncogene*. 2007; 26(24): 3451-3461.
- Beck BB, Baasner A, Buescher A, Habbig S, Reintjes N, Kemper M J, Sikora P, Mache C, Pohl M, Stahl M, *et al.* Novel findings in patients with primary hyperoxaluria type III and implications for advanced molecular testing strategies. *Eur J Hum Genet.* 2013; 21(2): 162-172.
- Bid HK, Kumar A, Kapoor R, Mittal RD. Association of Vitamin D Receptor-Gene (FokI) Polymorphism with Calcium Oxalate Nephrolithiasis. *J Endourol.* 2005; 19(1): 111-115.
- Bigelow MW, Wiessner JH, Kleinman JG, Mandel NS. Calcium oxalate-crystal membrane interactions: dependence on membrane lipid composition. J Urol. 1996; 155(3): 1094-1098.
- Bodmer W, Bonilla C. Common and rare variants in multifactorial susceptibility to common diseases. *Nat Genet.* 2008; 40(6): 695-701.
- Botstein D, Risch N. Discovering genotypes underlying human phenotypes: past successes for mendelian disease, future approaches for complex disease. *Nat Genet.* 2003; 33 Suppl: 228-237.
- Braun DA, Lawson JA, Gee HY, Halbritter J, Shril S, Tan W, Stein D, Wassner AJ, Ferguson MA, Gucev Z, *et al.* Prevalence of monogenic causes in pediatric patients with nephrolithiasis or nephrocalcinosis. *Clin J Am Soc Nephrol. 2016*; 11(4): 664-672.
- Brunette MG, Leclerc M. Renal action of progesterone: effect on calcium reabsorption. *Mol Cell Endocrinol.* 2002; 194(1-2): 183-190.
- Cellini B, Oppici E, Paiardini A, Montioli R. Molecular insights into primary hyperoxaluria type 1 pathogenesis. *Front Biosci (Landmark Ed)*. 2012; 17: 621-634.

- Chambers JC, Zhao J, Terracciano CM, Bezzina CR, Zhang W, Kaba R, Navaratnarajah M, Lotlikar A, Sehmi JS, Kooner MK, *et al.* Genetic variation in SCN10A influences cardiac conduction. *Nat Genet.* 2010; 42(2): 149-152.
- Chow K, Dixon J, Gilpin S, Kavanagh J P, Rao PN. Citrate inhibits growth of residual fragments in an in vitro model of calcium oxalate renal stones. *Kidney Int. 2004;* 65(5): 1724-1730.
- Coe FL, Evan A, Worcester E. Kidney stone disease. *J Clin Invest. 2005;* 115(10): 2598-2608.
- Coe FL, Parks JH, Moore ES. Familial idiopathic hypercalciuria. *N Engl J Med. 1979;* 300(7): 337-340.
- Cupisti A, Farnesi I, Armillotta N, Francesca F. Staghorn cystine stone in a 72-year-old recurrent calcium stone former. *Clin Nephrol.* 2012; 78(1): 76-80.
- Daga A, Majmundar AJ, Braun DA, Gee HY, Lawson JA, Shril S, Jobst-Schwan T, Vivante A, Schapiro D, Tan W, *et al.* Whole exome sequencing frequently detects a monogenic cause in early onset nephrolithiasis and nephrocalcinosis. *Kidney Int. 2018;* 93(1): 204-213.
- Danpure CJ, Rumsby G. Molecular aetiology of primary hyperoxaluria and its implications for clinical management. *Expert Rev Mol Med. 2004;* 6(1): 1-16.
- Dawn Teare M, Barrett J H. Genetic linkage studies. Lancet. 2005; 366(9490): 1036-1044.
- Ding Q, Fan B, Shi Y, Fan Z, Ding L, Li F, Tu W, Jin X, Qin C, Cao Q. Calcium-sensing receptor genetic polymorphisms and risk of developing nephrolithiasis in a Chinese population. *Urol Int.* 2017; 99(3): 331-337.
- Dressing GE, Thomas P. Identification of membrane progestin receptors in human breast cancer cell lines and biopsies and their potential involvement in breast cancer. *Steroids*. 2007; 72(2): 111-116.
- Evan AP, Coe FL, Lingeman JE, Worcester E. Insights on the pathology of kidney stone formation. *Urol Res.* 2005; 33(5): 383-389.
- Farber CR, Lusis AJ. Future of osteoporosis genetics: enhancing genome-wide association studies. *J Bone Miner Res. 2009;* 24(12): 1937-1942.
- Fernandes MS, Pierron V, Michalovich D, Astle S, Thornton S, Peltoketo H, Lam EW, Gellersen B, Huhtaniemi I, Allen J, *et al.* Regulated expression of putative membrane progestin receptor homologues in human endometrium and gestational tissues. J Endocrinol. 2005; 187(1): 89-101.
- Gambaro G, Vezzoli G, Casari G, Rampoldi L, D'Angelo A, Borghi L. Genetics of hypercalciuria

and calcium nephrolithiasis: From the rare monogenic to the common polygenic forms. *Am J Kidney Dis. 2004;* 44(6): 963-986.

- Gao B, Yasui T, Itoh Y, Li Z, Okada A, Tozawa K, Hayashi Y, Kohri K. Association of osteopontin gene haplotypes with nephrolithiasis. *Kidney Int.* 2007a; 72(5): 592-598.
- Gao B, Yasui T, Itoh Y, Tozawa K, Hayashi Y, Kohri K. A polymorphism of matrix Gla protein gene is associated with kidney stones. *J Urol. 2007b*; 177(6): 2361-2365.
- Garrison SR, Weyer AD, Barabas ME, Beutler BA, Stucky CL. A gain-of-function voltage-gated sodium channel 1.8 mutation drives intense hyperexcitability of A- and C-fiber neurons. *Pain.* 2014; 155(5): 896-905.
- Gaudet S, Branton D, Lue R A. Characterization of PDZ-binding kinase, a mitotic kinase. *Proc Natl Acad Sci U S A. 2000;* 97(10): 5167-5172.
- Gilissen C, Hoischen A, Brunner H G, Veltman J A. Unlocking Mendelian disease using exome sequencing. *Genome Biol.* 2011; 12(9): 228.
- Goldfarb DS, Fischer ME, Keich Y, Goldberg J. A twin study of genetic and dietary influences on nephrolithiasis: a report from the Vietnam Era Twin (VET) Registry. *Kidney Int. 2005;* 67(3): 1053-1061.
- Griffin DG. A review of the heritability of idiopathic nephrolithiasis. *J Clin Pathol.* 2004; 57(8): 793-796.
- Halbritter J. Genetics of kidney stone disease-Polygenic meets monogenic. *Nephrol Ther. 2021;* 17S: S88-S94.
- Halbritter J, Baum M, Hynes AM, Rice SJ, Thwaites DT, Gucev ZS, Fisher B, Spaneas L, Porath JD, Braun DA, *et al.* Fourteen monogenic genes account for 15% of nephrolithiasis/nephrocalcinosis. *J Am Soc Nephrol. 2015;* 26(3): 543-551.
- Hoopes RR, Jr., Shrimpton AE, Knohl SJ, Hueber P, Hoppe B, Matyus J, Simckes A, Tasic V, Toenshoff B, Suchy S F, *et al.* Dent Disease with mutations in OCRL1. *Am J Hum Genet.* 2005; 76(2): 260-267.
- Howles SA, Wiberg A, Goldsworthy M, Bayliss AL, Gluck AK, Ng M, Grout E, Tanikawa C, Kamatani Y, Terao C, *et al.* Genetic variants of calcium and vitamin D metabolism in kidney stone disease. *Nat Commun. 2019;* 10(1): 5175.
- Hu D, Barajas-Martinez H, Pfeiffer R, Dezi F, Pfeiffer J, Buch T, Betzenhauser MJ, Belardinelli L, Kahlig KM, Rajamani S, *et al.* Mutations in SCN10A are responsible for a large fraction of cases of Brugada syndrome. *J Am Coll Cardiol.* 2014; 64(1): 66-79.

- Johnson CM, Wilson DM, O'Fallon WM, Malek RS, Kurland LT. Renal stone epidemiology: a 25-year study in Rochester, Minnesota. *Kidney Int.* 1979; 16(5): 624-631.
- Khan SR. Reactive oxygen species as the molecular modulators of calcium oxalate kidney stone formation: evidence from clinical and experimental investigations. *J Urol. 2013;* 189(3): 803-811.
- Khaskhali MH, Byer K J, Khan S R. The effect of calcium on calcium oxalate monohydrate crystalinduced renal epithelial injury. *Urol Res. 2009;* 37(1): 1-6.
- Koyuncu HH, Yencilek F, Eryildirim B, Sarica K. Family history in stone disease: how important is it for the onset of the disease and the incidence of recurrence? *Urol Res. 2010;* 38(2): 105-109.
- Lai KC, Lin WY, Man KM, Tsai CH, Chen HY, Tsai FJ, Chen FJ, Chen HY, Liu HP, Ho TJ. Association of interleukin-18 gene polymorphisms with calcium oxalate kidney stone disease. *Scand J Urol Nephrol. 2010;* 44(1): 20-26.
- Li H, Zhang J, Long J, Shi J, Luo Y. Calcium-sensing receptor gene polymorphism (rs7652589) is associated with calcium nephrolithiasis in the population of Yi nationality in Southwestern China. *Ann Hum Genet. 2018a;* 82(5): 265-271.
- Li X, Dang X, Cheng Y, Zhang D, Zhang X, Zou T, Xing J. Common variants in ALPL gene contribute to the risk of kidney stones in the Han Chinese population. *Genet Test Mol Biomarkers*. 2018b; 22(3): 187-192.
- Lieske JC, Deganello S, Toback F G. Cell-crystal interactions and kidney stone formation. *Nephron*. *1999*; 81 Suppl 1: 8-17.
- Lieske JC, Rule AD, Krambeck AE, Williams JC, Bergstralh EJ, Mehta RA, Moyer TP. Stone composition as a function of age and sex. *Clin J Am Soc Nephrol. 2014;* 9(12): 2141-2146.
- Loredo-Osti JC, Roslin NM, Tessier J, Fujiwara TM, Morgan K, Bonnardeaux A. Segregation of urine calcium excretion in families ascertained for nephrolithiasis: evidence for a major gene. *Kidney Int. 2005;* 68(3): 966-971.
- Lu X, Sun D, Xu B, Pan J, Wei Y, Mao X, Yu D, Liu H, Gao B. In silico screening and molecular dynamic study of nonsynonymous single nucleotide polymorphisms associated with kidney stones in the SLC26A6 gene. J Urol. 2016; 196(1): 118-123.
- MacArthur J, Bowler E, Cerezo M, Gil L, Hall P, Hastings E, Junkins H, McMahon A, Milano A, Morales J, *et al.* The new NHGRI-EBI Catalog of

published genome-wide association studies (GWAS Catalog). *Nucleic Acids Res. 2017;* 45(D1): D896-D901.

- Madore F, Stampfer MJ, Rimm EB, Curhan GC. Nephrolithiasis and risk of hypertension. *Am J Hypertens.* 1998; 11(1 Pt 1): 46-53.
- Matsumoto S, Abe Y, Fujibuchi T, Takeuchi T, Kito K, Ueda N, Shigemoto K, Gyo K. Characterization of a MAPKK-like protein kinase TOPK. *Biochem Biophys Res Commun. 2004;* 325(3): 997-1004.
- Mesiano S, Wang Y, Norwitz ER. Progesterone receptors in the human pregnancy uterus: do they hold the key to birth timing? *Reprod Sci. 2011;* 18(1): 6-19.
- Mossetti G, Rendina D, Viceconti R, Manno G, Guadagno V, Strazzullo P, Nunziata V. The relationship of 3' vitamin D receptor haplotypes to urinary supersaturation of calcium oxalate salts and to age at onset and familial prevalence of nephrolithiasis. *Nephrol Dial Transplant. 2004;* 19(9): 2259-2265.
- National statistical office. 2021. Web site: http://www.nso.go.th (June,2021)
- Nettuwakul C, Praditsap O, Sawasdee N, Rungroj N, Ruamyod K, Watanapa WB, Junking M, Sangnual S, Sritippayawan S, Cheunsuchon B, *et al.* Lossof-function mutations of SCN10A encoding NaV1.8 alpha subunit of voltage-gated sodium channel in patients with human kidney stone disease. *Sci Rep. 2018;* 8(1): 10453.
- Nettuwakul C, Sawasdee N, Praditsap O, Rungroj N, Pasena A, Dechtawewat T, Deejai N, Sritippayawan S, Rojsatapong S, Chaowagul W, *et al.* A novel loss-of-function mutation of PBK associated with human kidney stone disease. *Sci Rep. 2020;* 10(1): 10282.
- Nicolaidou P, Themeli S, Karpathios T, Georgouli H, Athanassaki K, Xaidara A, Messaritakis J. Family pattern of idiopathic hypercalciuria and its subtypes. *J Urol*. *1996*; 155(3): 1042-1044.
- Nimmannit S, Malasit P, Susaengrat W, Ong-Aj-Yooth S, Vasuvattakul S, Pidetcha P, Shayakul C, Nilwarangkur S. Prevalence of endemic distal renal tubular acidosis and renal stone in the northeast of Thailand. *Nephron. 1996;* 72(4): 604-610.
- Pang Y, Dong J, Thomas P. Characterization, neurosteroid binding and brain distribution of human membrane progesterone receptors delta and {epsilon} (mPRdelta and mPR{epsilon}) and mPRdelta involvement in neurosteroid inhibition of apoptosis. *Endocrinology*. 2013; 154(1): 283-295.

- Pungsrinont T, Nettuwakul C, Sawasdee N, Rungroj N, Sritippayawan S, Yenchitsomanus P T. Association between intelectin-1 variation and human kidney stone disease in northeastern Thai population. Urolithiasis. 2021.
- Reed BY, Heller HJ, Gitomer WL, Pak CY. Mapping a gene defect in absorptive hypercalciuria to chromosome 1q23.3-q24. J Clin Endocrinol Metab. 1999; 84(11): 3907-3913.
- Romero V, Akpinar H, Assimos DG. Kidney stones: a global picture of prevalence, incidence, and associated risk factors. *Rev Urol.* 2010; 12(2-3): e86-96.
- Rule AD, Bergstralh EJ, Melton LJ, 3rd, Li X, Weaver AL, Lieske JC. Kidney stones and the risk for chronic kidney disease. *Clin J Am Soc Nephrol. 2009*; 4(4): 804-811.
- Rule AD, Roger VL, Melton LJ, 3rd, Bergstralh EJ, Li X, Peyser PA, Krambeck AE, Lieske JC. Kidney stones associate with increased risk for myocardial infarction. J Am Soc Nephrol. 2010; 21(10): 1641-1644.
- Rungroj N, Nettuwakul C, Sawasdee N, Sritippayawan S, Yenchitsomanus PT. Correlation between genotypes of F2 rs5896 (p.Thr165Met) polymorphism and urinary prothrombin fragment 1. *Urolithiasis*. 2018; 46(4): 405-407.
- Rungroj N, Nettuwakul C, Sudtachat N, Praditsap O, Sawasdee N, Sritippayawan S, Chuawattana D, Yenchitsomanus PT. A whole genome SNP genotyping by DNA microarray and candidate gene association study for kidney stone disease. BMC Med Genet. 2014; 15: 50.
- Rungroj N, Sritippayawan S, Thongnoppakhun W, Paemanee A, Sawasdee N, Nettuwakul C, Sudtachat N, Ungsupravate D, Praihirunkit P, Chuawattana D, *et al.* Prothrombin haplotype associated with kidney stone disease in Northeastern Thai patients. *Urology*. 2011; 77(1): 249 e217-223.
- Rungroj N, Sudtachat N, Nettuwakul C, Sawasdee N, Praditsap O, Jungtrakoon P, Sritippayawan S, Chuawattana D, Borvornpadungkitti S, Predanon C, et al. Association between human prothrombin variant (T165M) and kidney stone disease. PLoS One. 2012; 7(9): e45533-e45533.
- Saigal CS, Joyce G, Timilsina AR, Urologic Diseases in America P. Direct and indirect costs of nephrolithiasis in an employed population: opportunity for disease management? *Kidney Int.* 2005; 68(4): 1808-1814.
- Sayer JA. Progress in Understanding the Genetics of Calcium-Containing Nephrolithiasis. J Am Soc Nephrol. 2017; 28(3): 748-759.

- Scales CD, Jr., Smith AC, Hanley JM, Saigal CS, Urologic Diseases in America P. Prevalence of kidney stones in the United States. *Eur Urol.* 2012; 62(1): 160-165.
- Shughrue PJ, Stumpf WE, Sar M. The distribution of progesterone receptor in the 20-day-old fetal mouse: an autoradiographic study with [125I]progestin. *Endocrinology*. 1988; 123(5): 2382-2389.
- Sriboonlue P, Prasongwatana V, Chata K, Tungsanga K. Prevalence of upper urinary tract stone disease in a rural community of north-eastern Thailand. *Br J Urol.* 1992; 69(3): 240-244.
- Sriboonlue P, Prasongwatana V, Suwantrai S, Bovornpadungkitti S, Tungsanga K, Tosukhowong P. Nutritional potassium status of healthy adult males residing in the rural northeast Thailand. J Med Assoc Thai. 1998; 81(3): 223-232.
- Sriboonlue P, Prasongwattana V, Tungsanga K, Tosukhowong P, Phantumvanit P, Bejraputra O, Sitprija V. Blood and urinary aggregator and inhibitor composition in controls and renal-stone patients from northeastern Thailand. *Nephron.* 1991; 59(4): 591-596.
- Sritippayawan S, Borvornpadungkitti S, Paemanee A, Predanon C, Susaengrat W, Chuawattana D, Sawasdee N, Nakjang S, Pongtepaditep S, Nettuwakul C, *et al.* Evidence suggesting a genetic contribution to kidney stone in northeastern Thai population. *Urol Res. 2009;* 37(3): 141-146.
- Stapleton A, Ryall U. Blood coagulation proteins and urolithiasis are linked: crystal matrix protein is the Fl activation peptide of human prothrombin. *Br J Urol. 1995;* 75(6): 712-719.
- Stewart CS, Duncan SH, Cave DR. Oxalobacter formigenes and its role in oxalate metabolism in the human gut. *FEMS Microbiol Lett.* 2004; 230(1): 1-7.
- Suzuki K, Moriyama M, Nakajima C, Kawamura K, Miyazawa K, Tsugawa R, Kikuchi N, Nagata K. Isolation and partial characterization of crystal matrix protein as a potent inhibitor of calcium oxalate crystal aggregation: evidence of activation peptide of human prothrombin. Urol Res. 1994; 22(1): 45-50.
- Taguchi K, Yasui T, Milliner DS, Hoppe B, Chi T. Genetic risk factors for idiopathic urolithiasis: A systematic review of the literature and causal network analysis. *Eur Urol Focus. 2017;* 3(1): 72-81.
- Tang YT, Hu T, Arterburn M, Boyle B, Bright JM, Emtage PC, Funk W D. PAQR proteins: a novel membrane receptor family defined by an ancient 7-transmembrane pass motif. J Mol Evol. 2005; 61(3): 372-380.

- Tencza AL, Ichikawa S, Dang A, Kenagy D, McCarthy E, Econs MJ, Levine MA. Hypophosphatemic rickets with hypercalciuria due to mutation in SLC34A3/type IIc sodium-phosphate cotransporter: presentation as hypercalciuria and nephrolithiasis. *J Clin Endocrinol Metab.* 2009; 94(11): 4433-4438.
- Torricelli FC, Reichard C, Monga M. Urolithiasis in complicated inflammatory bowel disease: a comprehensive analysis of urine profile and stone composition. *Int Urol Nephrol.* 2021; 53(2): 205-209.
- Tosukhowong P, Borvonpadungkitti S, Prasongwatana V, Tungsanga K, Jutuporn S, Dissayabutr T, Reungjui S, Sriboonlue P. Urinary citrate excretion in patients with renal stone: roles of leucocyte ATP citrate lyase activity and potassium salts therapy. *Clin Chim Acta. 2002;* 325(1-2): 71-78.
- Trinchieri A, Ostini F, Nespoli R, Rovera F, Montanari E, Zanetti G. A prospective study of recurrence rate and risk factors for recurrence after a first renal stone. *J Urol.* 1999; 162(1): 27-30.
- Tsuji S, Uehori J, Matsumoto M, Suzuki Y, Matsuhisa A, Toyoshima K, Seya T. Human intelectin is a novel soluble lectin that recognizes galactofuranose in carbohydrate chains of bacterial cell wall. *J Biol Chem. 2001;* 276(26): 23456-23463.
- Udomsilp P, Saepoo S, Ittiwut R, Shotelersuk V, Dissayabutra T, Boonla C, Tosukhowong P. rs11567842 SNP in SLC13A2 gene associates with hypocitraturia in Thai patients with nephrolithiasis. *Genes Genomics.* 2018; 40(9): 965-972.
- Vezzoli G, Terranegra A, Rainone F, Arcidiacono T, Cozzolino M, Aloia A, Dogliotti E, Cusi D, Soldati L. Calcium-sensing receptor and calcium kidney stones. *J Transl Med. 2011;* 9: 201.

- Wang S, Wang X, Wu J, Lin Y, Chen H, Zheng X, Zhou C, Xie L. Association of vitamin D receptor gene polymorphism and calcium urolithiasis in the Chinese Han population. *Urol Res. 2012;* 40(4): 277-284.
- Wesener DA, Wangkanont K, McBride R, Song X, Kraft MB, Hodges HL, Zarling LC, Splain RA, Smith DF, Cummings RD, *et al.* Recognition of microbial glycans by human intelectin-1. *Nat Struct Mol Biol.* 2015; 22(8): 603-610.
- Wolf MT, Zalewski I, Martin FC, Ruf R, Muller D, Hennies HC, Schwarz S, Panther F, Attanasio M, Acosta HG, *et al.* Mapping a new suggestive gene locus for autosomal dominant nephrolithiasis to chromosome 9q33.2-q34.2 by total genome search for linkage. *Nephrol Dial Transplant. 2005;* 20(5): 909-914.
- Xu C, Song R-j, Yang J, Jiang B, Wang X-l, Wu W, Zhang W. Klotho gene polymorphism of rs3752472 is associated with the risk of urinary calculi in the population of Han nationality in Eastern China. *Gene. 2013;* 526(2): 494-497.
- Yanagawa M, Kawamura J, Onishi T, Soga N, Kameda K, Sriboonlue P, Prasongwattana V, Borwornpadungkitti S. Incidence of urolithiasis in northeast Thailand. *Int J Urol. 1997*; 4(6): 537-540.
- Yasui T, Okada A, Urabe Y, Usami M, Mizuno K, Kubota Y, Tozawa K, Sasaki S, Higashi Y, Sato Y. A replication study for three nephrolithiasis loci at 5q35. 3, 7p14. 3 and 13q14. 1 in the Japanese population. *J Hum Genet.* 2013; 58(9): 588-593.