Isolation and identification of *Aspergillus* species producing Ochratoxin A in Arabica coffee beans

Sumanee Kuntawee and Angsana Akarapisan*

Department of Entomology and Plant Pathology, Faculty of Agriculture, Chiang Mai University, Chiang Mai 50200, Thailand

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Isolation of fungi in Arabica parchment coffee and green coffee were collected from in Omkoi district, Chiang Mai, Thailand. The coffee beans were isolated by direct plating method. The postharvest fungi of Arabica parchment coffee and green coffee were assessed for the presence of *Aspergillus* species, the ochratoxigenic potential of the isolates and ochratoxin A levels. Contamination by *Aspergillus* and *Rhizopus* species was found on 25.46% and 38.88% respectively, of 12 samples. Sixteen isolates were identified to species comprised *Aspergillus* section *Ochraceus* (3), *Ostianus* (2), *Candidus* (2), *Sclerotiorum*, *Awamori*, *Parasiticus*, *Terreus* and 5 isolated of *Aspergillus* spp. not subsequently species. The fungi group of *Aspergillus* were isolated to preliminary determine the Ochratoxin A (OTA) on Coconut agar (CA) incubated at 28 C° for 7 day, then observed the fluorescence under UV light with a wavelength of 365 nm. The result showed that 12 isolates of *Aspergillus* species produced fluorescence blue on CA medium. Confirmed by High Performance Liquid Chromatography (HPLC) found that 10 isolates produced OTA in the range of 0.004 to 12,937.50 ng/g. *Aspergillus ochraceus* isolates produced the maximum OTA observed in agar with toxigenic species.

Keywords: Ochratoxin A (OTA), Arabica coffee beans, Fluorescence, *Aspergillus* group, HPLC.

Introduction

The importance of coffee in the world economy, coffee growing and drinking spread around the world. The most important strain of coffee

^{.*}Corresponding author: Angsana Akarapisan; e-mail: angsana.a@cmu.ac.th

economically are Coffee Arabica L. which accounts for over 70% of global production. The Ochratoxin A (OTA) has been reported from temperate to tropical climates mainly on coffee and their products. The OTA a nephrotoxic and possible carcinogen has been detected in a number of foods. In the experiment was done by Batista et al. (2003), processed (green) coffee beans in Brazil were assessed both before and after surface sterilization contamination by Aspergillus and Penicillium species was found on 96% and 42% respectively. This is in agreement with data presented by Noomim et al. (2008) Aspergillus steynii (13/13) was the best ochratoxin producer, because all strains consistently produced both OTA and OTB in large amounts. Isolates of Aspergillus westerdijkiae (42/42) were also consistent and 100% of them produced ochratoxin. There are also reports by Frisrad et al. (2004) the fungus Yellow Aspergillus sp. have been produce of Ochratoxin A toxin such as Aspergillus cretensis, Aspergillus flocculosus, Aspergillus neobridgeri, Aspergillus steynii, Aspergillus pseudoelegans, Aspergillus roseoglobulosus and Aspergillus westerdijkiae. Robusta coffee beans were infected by fungi more than Arabica coffee. Aspergillus niger infected 89% of Robusta beans, whereas Aspergillus carbonarius and yellow Aspergilli each infected 12-14% of beans. The OTA was not produced by A. niger (98 isolates) or A. ochraceus (77 isolates), but was detected in 110 of 113 isolates of A. carbonarius, 10 isolates of A. westerdijkiae and one isolate of A. steynii (Leong et al., 2007). Postharvest diseases may significantly lower the quality and quantity of this produces, most postharvest fungi on chili are storage fungi such as the fungal genera Aspergillus, Penicillium, Rhizopus, Cladosporium and Trichoderma (Suwan and Akarapisan, 2012). The reported incidence of toxigenicity among strains of A. carbonarius isolated from grapes ranges between 58% and 97% of strains surveyed (Battilani et al., 2003) The objective of study was to investigate the distribution of fungi with the incidence and toxigenicity of OTA-producing Aspergillus species infecting Arabica parchment coffee.

Materials and Methods

Isolation of fungi from Arabica coffee beans

The first step was performed using the method of Leong *et al.* (2007) as follows. The Arabica coffee beans were collected from Omkoi district, Chiang Mai. The Postharvest fungi were isolated by direct plating method. Fifty beans per sample were plated on water agar (WA) (10 beans/plate) incubated at 28 C^o for 7 days. Fungi were identified under the stereomicroscope.

Preliminary screening with Ochratoxin A (OTA) on Coconut agar

The isolates fungi group of *Aspergillus* were single point inoculated on Coconut agar (CA) incubated at 28 C° for 3 days in the dark. To detect fluorescence, the reverse side of the CA plates was viewed under UV light with a wavelength of 365 nm. An uninoculated CA plate was used as a control. Species that do not produce OTA were also inoculated on CA plates as negative controls.

Determination Ochratoxin A (OTA) of fungi by HPLC

All of identification the isolates of *Aspergillus* group which produced OTA were inoculated in Yeast Extract Sucrose (YES) agar. The strains were three-point in YES agar incubated at 28 $^{\circ}$ for 7 day in the dark. Three agar plugs were removed from area of the colony, 1 g of YES agar were added in 2 ml H₂O beaker and mashed homogenization. Then, the 1ml solutions were filtered through a syringe filter into a brown bottle. The injection volume was 100 μ l onto the HPLC (model SCL-10AVP SHIMADZU).

Identification of Aspergillus group

All isolate of *Aspergillus* group which produced OTA were analyzed species *Aspergillus* by using Biolog FF microplate. (Prima Scientific Co.,Ltd)

Results and discussions

Fungal infection and Identification of fungi isolates

Infection with one or more fungi, with Yellow Aspergillus and Black Aspergillus being the dominate species, disease incidence of fungi on Arabica parchment coffee (CNK1a) and green coffee (CNK1b) showed that percent infection with 63.9% and 45.8%. The contaminated with Aspergillus species and Rhizopus sp. respectively. For isolate Arabica parchment coffee (CNK2) disease incidence of fungi showed percent infection with 69.4% and isolate Arabica parchment coffee (CNK3) disease incidence of fungi showed the highest percent infection with 100%. Perrone et al. (2003) reported that Arabica coffee bean samples from the North Thailand, two types of samples, parchment coffee bean and green coffee beans were showed that approximately

75 % of the samples were contaminated by black aspergilli and similar levels of contamination were observed for isolates belonging to *Aspergillus* section Circumdati. As well as Taniwaki *et al.* (2003) reported that *A. ochraceus* was the dominant yellow *Aspergillus* species in Brazilinan coffee.

Contaminating coffee samples were isolated and identified. Aspergillus specie producers Ochratoxin A (OTA) in Yellow Aspergillus, Black Aspergillus and Green Aspergillus Groups. Aspergillus species were subsequently identified by Biolog, as three isolates of Aspergillus ochraceus, two isolates of Aspergillus ostianus, two isolates of Aspergillus candidus, one isolates of Aspergillus sclerotiorum, one isolates of Aspergillus awamori, one isolates of Aspergillus parasiticus, one isolates of Aspergillus terreus. Five isolates of Aspergillus sp. was not subsequently identified to species. Noonim et al. (2003) reported that A. sclerotiicarbonarius is related to A. carbonarius and A. ibericus and was found only in the Southern region of Thailand. The A. ochraceus and A. ostianus were two predominant species, comprising 18.75% and 12.5% on Arabica parchment coffee. Morello et al. (2007) reported that by using sequences of the β-tubulin gene, which most isolates from Brazilian coffee (84%) previously identified as Aspergillus ochraceus were actually A. westerdijkiae.

Screening of Aspergillus isolates for Ochratoxin A (OTA) on Coconut agar

A simple and rapid screening method was developed for the detection of OTA in fungi cultures using Coconut agar (CA). Three groups of *Aspergillus* as yellow, black and green were tested preliminary determine produced OTA on CA. The plates were incubated at 28 C° for 3 days then, examined for pigmentation and fluorescence under UV light (365 nm). Twelve isolates of Yellow *Aspergillus* which, OTA produces a blue fluorescence usually covering the whole colony on CA. *Aspergillus parasiticus* produced a yellow green fluorescence usually covering the colony on CA not produced OTA (Fig. 1a) and *Aspergillus* sp. (Yellow) which, OTA produces a blue fluorescence medium on CA (Fig. 1b). *Aspergillus ochraceus* which, OTA produced a blue fluorescence strong on CA (Fig. 1c) and *Aspergillus terreus* that did not produce pigmentation a blue fluorescence on CA were considered to be OTA-negative (Fig. 1d). Ochratoxigenic *Aspergillus ochraceus* and *Aspergillus ostianus* produced OTA. Heenan *et al.* (1998) previous reported OTA produced

a blue green fluorescence usually covering the colony on coconut cream agar. As well as Mantle and Chow (2000) reported that Asian coffee isolates produced ochratoxin A the range 400 mg kg-1, on coconut agar OTA appears as an intense blue fluorescence.

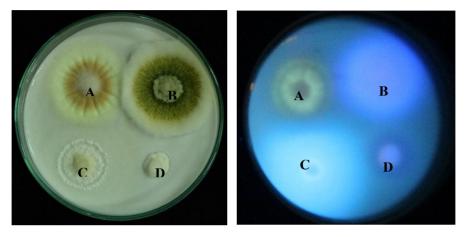


Fig. 1. Ohratoxin A production by *Aspergillus* groups culture on Coconut Agar (CA) (A) *Aspergillus parasiticus*, (B) *Aspergillus* sp. (Yellow) (C) *Aspergillus ochraceus* and (D). *Aspergillus terreus*, fig right viewed under long wavelength UV (365 nm).

HPLC analysis

Three groups of *Aspergillus* as yellow, black and green were tested preliminary determine produced OTA on CA. The result showed that 12 isolates of *Aspergillus* species produced fluorescence blue on CA medium. In confirmed produced OTA using the agar plug HPLC method on Yeast Extract Sucrose (YES) agar. Twelve isolates of *Aspergillus* were detected for OTA production. The *Aspergillus ochraceus* was produced OTA levels higher than 12,937.50 ng/g (Table 1). Chen *et al.* (2013) reported that *Penicillium chrysogenum*, *Penicillium glycyrrhizacola*, *Penicillium polonicum*, *Aspergillus ochraceus* and *Aspergillus westerdijkiae* could produce OTA, concentration varied among the isolates from 12.99 to 39.03 µg/kg. Similar work has reported high Ochratoxin A production (400–16,000 µg/kg) in grains such as wheat, rye and barley, showing that some isolates of *Aspergillus ochraceus* are high producers of this toxin (Kononenko *et al.*, 2000).

Table 1. Detection of Ochratoxin A (OTA) by Coconut agar (CA), Potato dextrose agar (PDA), Yeast extract sucrose agar (YES) direct method and by the HPLC agar plug method

Isolate	CA ¹	PDA ²	YES ³	HPLC (ng/g)	Biolog
Cnk2/a	B (+)	-	-	1651.34	Aspergillus sp. (yellow)
Cnk2/b	B (+++)	B (+)	-	152.42	Aspergillus ostianus
Cnk2/c	B (++)	B (+)	-	4122.44	Aspergillus sclerotiorum
Cnk3/a	B (++)	-	-	190.32	Aspergillus ochraceus
Cnk3/b	B (+++)	B (+)	-	4202.32	Aspergillus ochraceus
Cnk3/c	B (+++)	B (+)	-	6900.90	Aspergillus ostianus
Cnk3/d	+	-	-	269.28	Aspergillus sp. (yellow)
Cnk3/e	B (++)	-	-	583.12	Aspergillus sp. (yellow)
Cnk3/f	G (+)	-	-	nd	Aspergillus parasiticus
Cnk3/g	B (++)	B (+)	-	2627.02	Aspergillus candidus
Cnk3/h	B (+++)	B (++)	B (+)	12,937.5	Aspergillus ochraceus
Cnk3/i	B (+++)	B (+)	-	8,972.16	Aspergillus candidus
(Negative control)	-	-	-	nd	Aspergillus terreus
(Negative control)	-	-	-	nd	Aspergillus sp. (yellow)
D/2 (green Aspergillus)	B (+)	-	-	0.014	Aspergillus sp. (green)
N/2 (black Aspergillus)	B (+)	-	-	0.004	Aspergillus awamori
¹ Coconut agar	² Potato devtrose agar			³ Veast extract sucrose	

¹Coconut agar

²Potato dextrose agar

³Yeast extract sucrose

B (+++): Create a fluorescence blue strong B (++): Create a fluorescence blue medium

B (+): Create a fluorescence blue weak G (+): Create fluorescence yellow-green. (-): do not fluorescence.

nd: not detected

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References

- Batista, R.L., Chalfoun, M.S., Prado, G., Schwan, R.F. and Wheals, A.E. 2003. Toxigenic fungi associated with processed (green) coffee beans (*Coffea arabica* L.). Journal of Food Microbiology 85: 293 300.
- Battilani, P., Pietri, A., Bertuzzi, T., Languasco, L., Giorni, P. and Kozakiewicz, Z. 2003. Occurrence of ochratoxin A-producing fungi in grapes grown in Italy. Journal of Food Protection 66: 633–636.
- Chen, A. J., Tang, D., Zhou, Y. Q., Sun, B. D., Li, X. J., Wang, L. Z. and Gao, W. W. 2013. Identification of Ochratoxin A Producing Fungi Associated with Fresh and Dry Liquorice. Public Library of Science 8: 1-13.
- Frisvad, C. J., Frank, M. J., Houbraken, A.M.P. J., Kuijpers, F.A. A. and Samson, A. R. 2004. New ochratoxin a producing species of *Aspergillus* section Circumdati. Studies in mycology 50: 23–43.
- Heenan, C.N., Shaw, K.J. and Pitt, J.I. (1998) Ochratoxin A production by *Aspergillus carbonarius* and *Aspergillus niger* isolates and detection using conconut cream agar. Journal of Food Microbiology 1: 67–72.
- Kononenko, G. P., Burkin, A. A., Zotova, E. V. and Soboleva, N. A. (2000). Ochratoxin A: contamination of grain. Applied Biochemistry and Microbiology 36: 177– 180.
- Leong, S. L., Hien, L. T., An, T. V., Trang, N. T., Hocking, A. D. and Scott, E. S. 2007. Ochratoxin A- producing Aspergilli in Vietnamese green coffee beans. Letters in Applied Microbiology 45: 301–306.
- Mantle, G. P. and Chow, M. A. 2000. Ochratoxin formation in *Aspergillus ochraceus* with particular reference to spoilage of coffee. International Journal of Food Microbiology 56: 105–109.
- Morello, L.G., Sartori, D., de Oliveira Martinez, A.L., Vieira, M.L.C., Taniwaki, M.H. and Fungaro, M.H.P. 2007. Detection and quantification of *Aspergillus westerdijkiae* in coffee beans based on selective amplification of β tubulin gene by using real-time PCR. International Journal of Food Microbiology 119: 270–276.
- Noonim, P., Mahakarnchanakul W., Nielsen K.F., Frisvad J.C. and Samson R.A. 2009. Fumonisin B2 production by *Aspergillus niger* in Thai coffee beans. Food Additives and Contaminants 26: 94–100.
- Perrone, G., Suscal, A., Cozzi, G., Ehrlich, K., Varga, J. Frisvad, J.C. Meijer, M. Noonim, P., Mahakarnchanakul, W. and Samson, R. A. 2007. Biodiversity of

- Aspergillus species in some important agricultural products. Studies in Mycology 59: 53–66.
- Suwan, N. and Akarapisan, A. 2012. Screening of Antagonistic Bacteria to Control Postharvest Disease of Karen Chili. Paper presented in the 1st ASEAN Plus Three Graduate Research Congress (AGRC 2012). 323-327 p.
- Taniwaki, M.H., Pitt, J.I., Teixeira, A.A. and Iamanaka, B.T. 2003. The source of ochratoxin A in Brazilian coffee and its formation in relation to processing methods. International Journal of Food Microbiology 82: 173–179.