



The Genus *Phaeosaccardinula* (Chaetothyriales) from Yunnan, China, Introducing Two New Species

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ABSTRACT

This paper deals with two new species of *Phaeosaccardinula* collected in Yunnan Province of China. *Phaeosaccardinula multiseptata* and *P. dendrocalami* are introduced as new species, based on morphology and molecular data and are compared with related taxa. Descriptions, illustrations and notes are provided for each species. A phylogenetic tree based on analysis of ITS and LSU sequence data show the species to cluster in *Phaeosaccardinula* (Chaetothyriaceae) as two distinct clusters. Morphology and analysis of ITS, LSU and EF1a gene data indicate several differences between *P. ficus* and *P. multiseptata* in the first cluster, while the second cluster represents the new species *P. dendrocalami*. Previously, sequence data (ITS and LSU) were only available for *Phaeosaccardinula* species, this paper provides new sequence data for ITS, LSU, EF1a and RPB2 genes.

Key words: Chaetothyriaceae, EF1a, foliar epiphyte, ITS, LSU, phylogeny

1. INTRODUCTION

The *Chaetothyriaceae* was introduced by Hansford in 1946 [1]. The majority of species in this family are superficial on leaves and have no connexion with insects and their secretions [1]. However the ecology of many species is poorly studied and it is unclear whether they are saprotrophic or biotrophic [2]. The mycelia form a dense matt of hyphae, which have the appearance of sooty moulds and are appressed to the host cuticle without penetrating host tissues [3]. The *Chaetothyriaceae* includes 13 genera, with only *Ceramothyrium*, *Phaeosaccardinula* and *Yatesula* lacking setae [4].

Phaeosaccardinula has an estimated 14 species [5] which are foliar epiphytes, and have superficial ascocarps, with a dark, non-setose pellicle, saccate, bitunicate ascii and muriform, hyaline to brownish ascospores [3,6]. More than 40 species have epithets under this genus (Index Fungorum 2013), but many actually belong in *Limacinula* and *Treibiomycetes* [7,8]. A new species, *Phaeosaccardinula ficus* Chomnumti & K.D. Hyde, was introduced from northern Thailand by Chomnumti *et al.* [9]. This species with lacks a tessellate scutellum and spores are olivaceous green at the septa and have a

mucilaginous sheath [9].

Phaeosaccardinula javanica (Zimm.) Yamam., *P. longispora* Yamam. and *P. dictyospora* (Petr.) O. Erikss. have been reported from China [7,8,10]. However *P. javanica* is now named *Limacinula javanica* (Zimm.) Höhn. and *P. longispora* Yamam. is *Trebiomyces roseosporus* (Höhn.) D.R. Reynolds [10]. Therefore *P. dictyospora* is the only species in the *Phaeosaccardinula* known from China [10]. The species are listed as *Chaetothyrium javanicum* (Zimm.) Boedijn and *Trebiomyces roseosporus* (= *p. longispora* Yamam.) in Index Fungorum (2013), indicating differences in opinion and need for verification with molecular data. *Chaetothyrium javanicum* has been reported from at least 73 different hosts and surely must represent more than one *Limacinula* species [10].

The purpose of this study is to introduce two new species in the genus *Phaeosaccardinula* and we also add ITS, LSU, EF1a and RPB2 gene sequence data for this poorly known genus.

2. MATERIALS AND METHODS

2.1 Isolates and Morphology

Fungi with sooty mould-like colonization were collected from various living plants in Yunnan. Specimens were taken to the laboratory in an envelope and examined under a microscope (Nikon 80i) for morphological characters. Sections of ascocarps were made in a freezing microtome (Leica CM1100). Pure-cultures were obtained by single spore isolation following the methods of Chomnunti *et al.* [11]. After a month, the growing cultures were used for molecular work.

The specimens were air dried and are deposited at the International Fungal Research & Development Centre (IFRD) Herbarium, Kunming, Yunnan, and the cultures are stored in the culture collection of the same institution (IFRDCC) with duplicates in MFLUCC and

ICMP.

DNA isolation, amplification and sequencing: Fungal isolates were grown on PDA for 40 days at 26°C in the dark. Genomic DNA was extracted from the growing mycelium using the Biospin Fungus Genomic DNA Extraction Kit (BioFlux®) following the manufacturer's protocol (Hangzhou, P.R. China).

Polymerase chain reaction (PCR) was carried out using known primer pairs LROR and LR5 were used to amplify a region spanning the large subunit rDNA (28S, LSU) [12] and internal transcribed spacers (5.8S, ITS) was amplified by primer pairs ITS5 and ITS4 [13]. The translation elongation factor 1-alpha gene (EF1a) was amplified by using EF1-983F and EF1-2218R primers [14]. The RNA polymerase II second largest subunit (RPB2) was amplified by using RPB2-5F and RPB2-7cR primers [15]. The amplification reaction mixtures were 50 µL which contained 3.0 µL of DNA template, 1.5 µL of each forward and reverse primers, 25 µL of 2 × Easy Taq PCR SuperMix (mixture of EasyTaq™ DNA Polymerase, dNTPs, and optimized buffer, Beijing TransGen Biotech Co., Ltd., Chaoyang District, Beijing, PR China) and 19 µL sterilized water. Amplification conditions were set up for initial denaturation of 3 min at 94°C, followed by 38 cycles of 30 s at 94°C, 40 s at 56°C and 60 s at 72°C, and a final extension period of 10 min at 72°C [16]. The PCR products were observed on 1% agarose electrophoresis gels stained with ethidium bromide. Purification and sequencing of PCR products were carried at Shanghai Sangon Biological Engineering Technology & Services Co., Ltd (China).

2.2 Phylogenetic Analysis

Blast searches were made to reveal the closest matches in GenBank. All ITS and LSU sequences obtained from GenBank (listed in

Table 1.) [17-22]. The EF1a sequence for *Phaeosaccardinula ficus* was provided by P. Chomnunti (GenBank No. = KF791915). Sequences were aligned and concatenated in BioEdit [23]. Phylogenetic analyses were carried by using RAxML v. 7.2.7 [24] implemented in RAxMLGui v 1.3 [25] for Maximum likelihood (ML) and MrBayes v. 3.0b4 [26] for Bayesian analyses.

Maximum likelihood (ML) trees searches were done in RAxMLGui v 1.3 [24] and used the 'ML + thorough bootstrap' option. One thousand non parametric bootstrap iterations were run with the general time reversible (GTR) model and a discrete gamma distribution. The resulting replicates were plotted on to the best scoring tree obtained previously. The phylogram with bootstrap values above the branches is presented in

Figure 1. by using graphical options available in TreeView [27].

The model of evolution was performed by using MrModeltest v. 3.0b4. Posterior probabilities (PP) were determined by Markov Chain Monte Carlo sampling (BMCMC) in MrBayes v. 3.0b4 [28]. Six simultaneous Markov chains were run for 1,000,000 generations and trees were sampled every 100th generation and 10,000 trees were obtained. The first 2,000 trees, representing the burn-in phase of the analyses, were discarded while remaining 8,000 trees used for calculating posterior probabilities in the majority rule consensus tree [29,30]. Bayesian Posterior Probabilities (BYPP) with those equal or greater than 0.80 given below each node (Figure 1).

Table 1. Taxa used in the phylogenetic analysis and their corresponding GenBank numbers.

Species	Culture/voucher	ITS	LSU
<i>Antennariella placitae</i>	CBS 124785	GQ303268	GQ303299
<i>Capnodium coffeeae</i>	CBS 147.52	AJ244239	—
<i>Caproniamansorii</i>	—	AF050247	AY004338
<i>Caproniamumkii</i>	DAOM 216390	AF050250	EF413604
<i>Caproniasemimmersa</i>	MUCL 40572	AF050259	AF050259
<i>Ceramothyrium carniolicum</i>	CBS 175.95	KC978733	FJ358232
<i>Ceramothyrium thailandicum</i>	MFLU 10-0079	HQ895838	HQ895835
<i>Chaetothyrium brischoficola</i>	MFLU 10-0083	HQ895839	HQ895836
<i>Cladophialophora australiensis</i>	CBS 112793	EU035402	EU035402
<i>Cladophialophora minourae</i>	CBS 556.83	AY251087	FJ358235
<i>Cladophialophora potulentorum</i>	CBS 112222	EU035409	EU035409
<i>Conidioxyphium gardneriarum</i>	CPC 14327	—	GU301807
<i>Cyphellophora laciniata</i>	CBS 190.61	EU035416	FJ358239
<i>Cyphellophora pauciseptata</i>	CBS 284.85	JQ766469	JQ766522
<i>Cyphellophora pauciseptata</i>	CBS 284.85	JQ766470	JQ766523
<i>Cyphellophora suttonii</i>	CBS 449.91	GU225946	—
<i>Cyphellophora vermispora</i>	CBS 227.86	GU225947	JQ766474
<i>Exophiala pisciphila</i>	AFTOL-ID 669	DQ826739	DQ823101
<i>Exophiala salmonis</i>	CBS 157.67	JF747138	AY213702
<i>Leptoxyphium madagascariense</i>	CBS 124766	GQ303277	GQ303308
<i>Microxyphium theae</i>	CBS 202.30	— GU301849	
<i>Phaeosaccardinula dendrocalami</i>	IFRDCC 2649	KF667242	KF667245

Table 1. Continued.

Species	Culture/voucher	ITS	LSU
<i>Phaeosaccardinula dendrocalami</i>	IFRDCC 2663	KF667243	KF667246
<i>Phaeosaccardinula ficus</i>	MFLU 10-0009	HQ895840	HQ895837
<i>Phaeosaccardinula multiseptata</i>	IFRDCC 2639	KF667241	KF667244
<i>Placiopsis cinerascens</i>	MA 161308	GQ344613	GQ344570
<i>Placiopsis tenella</i>	L 1281352	GQ344616	GQ344575
<i>Placiopsis tirolensis</i>	L 1281349	GQ344618	GQ344581
<i>Rhinocladiella anceps</i>	CBS 157.54	EU041804	EU041861
<i>Rhinocladiella fasciculata</i>	CBS 132.86	EU041807	EU041864
<i>Trichomerium deniquatum</i>	MFLU 10-0884	JX313654	JX313660
<i>Trichomerium foliicola</i>	MFLU 10-0054	JX313651	JX313657
<i>Trichomerium foliicola</i>	MFLU 10-0058	JX313653	JX313659
<i>Trichomerium foliicola</i>	MFLU 10-0073	JX313652	JX313658
<i>Trichomerium gloesporum</i>	MFLU 10-0087	JX313656	JX313662
<i>Veronaea botryosa</i>	CBS 350.65	EU041817	EU041874
<i>Venturia inaequalis</i>	CBS 476.61	EU282478	GU456336

2.3 Individual Data for ITS, LSU, EF1a and RPB2 for Four Strains

Seqman in DNASTar software v5.01 (www.dnastar.com) [31] was used to compare

the different base pairs of ITS, LSU, EF1a and RPB2 between *P. ficus* and *P. multiseptata* and two strains of *P. dendrocalami*, the details of the difference are listed in Table 2.

Table 2. ITS, LSU, EF1a and RPB2 base pair difference between *P. ficus* and *P. multiseptata* and two strains of *P. dendrocalami*.

Species name	Gene name	Different bases number	Details of difference					
<i>P. dendrocalami</i> (IFRDCC2649 and IFRDCC2663)	ITS	1	<p style="text-align: center;">190</p> <table border="0"> <tr> <td style="vertical-align: top;"> <u>Translate ► Consensus</u> <u>IFRDCC2649.seq(1>520) →</u> <u>IFRDCC2663.seq(1>426) →</u> </td> <td style="vertical-align: top;"> <u>AAYTCTG</u> <u>AACTCTG</u> <u>AATTCTG</u> </td> </tr> </table>	<u>Translate ► Consensus</u> <u>IFRDCC2649.seq(1>520) →</u> <u>IFRDCC2663.seq(1>426) →</u>	<u>AAYTCTG</u> <u>AACTCTG</u> <u>AATTCTG</u>			
<u>Translate ► Consensus</u> <u>IFRDCC2649.seq(1>520) →</u> <u>IFRDCC2663.seq(1>426) →</u>	<u>AAYTCTG</u> <u>AACTCTG</u> <u>AATTCTG</u>							
	LSU	1	<p style="text-align: center;">660</p> <table border="0"> <tr> <td style="vertical-align: top;"> <u>Translate ► Consensus</u> <u>IFRDCC2649.seq(1>855) →</u> <u>IFRDCC2663.seq(1>1237) →</u> </td> <td style="vertical-align: top;"> <u>CCYCGGGCT</u> <u>CCCCGGGCT</u> <u>CCTCGGGCT</u> </td> </tr> </table>	<u>Translate ► Consensus</u> <u>IFRDCC2649.seq(1>855) →</u> <u>IFRDCC2663.seq(1>1237) →</u>	<u>CCYCGGGCT</u> <u>CCCCGGGCT</u> <u>CCTCGGGCT</u>			
<u>Translate ► Consensus</u> <u>IFRDCC2649.seq(1>855) →</u> <u>IFRDCC2663.seq(1>1237) →</u>	<u>CCYCGGGCT</u> <u>CCCCGGGCT</u> <u>CCTCGGGCT</u>							
	EF1a	4	<p style="text-align: center;">110</p> <table border="0"> <tr> <td style="vertical-align: top;"> <u>Translate ► Consensus</u> <u>IFRDCC2649.seq(1>931) →</u> <u>IFRDCC2663.seq(1>858) →</u> </td> <td style="vertical-align: top;"> <u>GTYACTTA</u> <u>GTTACTTA</u> <u>GTCACTTA</u> </td> <td style="vertical-align: top;"> <u>CCRGT</u> <u>CCAGT</u> <u>CCGGT</u> </td> </tr> </table> <p style="text-align: center;">230</p> <table border="0"> <tr> <td style="vertical-align: top;"> <u>GACTGGYG</u> <u>GACTGGCG</u> <u>GACTGGTG</u> </td> </tr> </table> <p style="text-align: center;">420</p> <table border="0"> <tr> <td style="vertical-align: top;"> <u>CTCGTGTRTG</u> <u>CTCGTGAATG</u> <u>CTCGTGGTG</u> </td> </tr> </table>	<u>Translate ► Consensus</u> <u>IFRDCC2649.seq(1>931) →</u> <u>IFRDCC2663.seq(1>858) →</u>	<u>GTYACTTA</u> <u>GTTACTTA</u> <u>GTCACTTA</u>	<u>CCRGT</u> <u>CCAGT</u> <u>CCGGT</u>	<u>GACTGGYG</u> <u>GACTGGCG</u> <u>GACTGGTG</u>	<u>CTCGTGTRTG</u> <u>CTCGTGAATG</u> <u>CTCGTGGTG</u>
<u>Translate ► Consensus</u> <u>IFRDCC2649.seq(1>931) →</u> <u>IFRDCC2663.seq(1>858) →</u>	<u>GTYACTTA</u> <u>GTTACTTA</u> <u>GTCACTTA</u>	<u>CCRGT</u> <u>CCAGT</u> <u>CCGGT</u>						
<u>GACTGGYG</u> <u>GACTGGCG</u> <u>GACTGGTG</u>								
<u>CTCGTGTRTG</u> <u>CTCGTGAATG</u> <u>CTCGTGGTG</u>								

Table 2. Continued.

Species name	Gene name	Different bases number	Details of difference
	RPB2	13	<p style="text-align: center;">160 250</p> <p>Translate ▶ Consensus CGCRTC ARCCAAAAGGTGGRT</p> <p>IFRDCC2649.seq(1>994) → CGCGTC AGCCAAAAGGTGGAT</p> <p>IFRDCC2663.seq(1>972) → CGCATC AACCCAAAAGGTGGT</p> <p style="text-align: center;">270 340 350 490 570 600</p> <p>AAYGG AGRTT TTGGCRC GTCTRA TGSG CYTCCAAC</p> <p>AATGG AGGTT TTGGCAC GTCTGA TGGG CTTCCAAC</p> <p>AACGG AGATT TTGGCGC GTCTAA TCCG CCTCCAAC</p> <p style="text-align: center;">640 680 750 890</p> <p>ARGGGCC AAATTYT TRITG TCTGRT</p> <p>AAGGGCC AAATTIT TATTG TCTGAT</p> <p>AGGGGCC AAATTCT TGTTG TCTGGT</p>
<i>P.ficus</i> (MFLUCC10-0009) and <i>P.multiseptata</i> (IFRDCC2639)	ITS	4	<p style="text-align: center;">100 110</p> <p>Translate ▶ Consensus GWGAGGGTCTCCGRG</p> <p>MFLUCC10-0009.seq(1>627) → GAGAGGGTCTCCGGG</p> <p>IFRDCC2639.seq(1>618) → GTGAGGGTCTCCGAG</p> <p style="text-align: center;">140 200</p> <p>TACTSTG CGCCCCGYC</p> <p>TACTCTG CGCCCGCC</p> <p>TACTGTG CGCCCGTC</p>
	LSU	6	<p style="text-align: center;">530 900</p> <p>Translate ▶ Consensus CCWCCC AGKKGGTT</p> <p>MFLUCC10-0009.seq(1>1474) → CCTCCC AGTGGGTT</p> <p>IFRDCC2639.seq(1>880) → CCAACC AGGTGGTT</p> <p style="text-align: center;">1140 1150</p> <p>TKGCATAKGGTGGGGRC</p> <p>TTGCATTGGTGGGGGAC</p> <p>TGGCAGTGGTGGGGGCG</p>
	EF1a	1	<p style="text-align: center;">160</p> <p>Translate ▶ Consensus GTGRGC</p> <p>MFLUCC10-0009.seq(1>920) → GTGAGC</p> <p>IFRDCC2639.seq(1>860) → GTGGGC</p>

Note: The different base pairs of the sequence are marked in red. The upper number is the placement of the base pairs. *Phaeosaccardinula ficus* = MFLUCC 10-0009, *P. multiseptata* = IFRDCC 2639, *P. dendrocalami* = IFRDCC 2649 and IFRDCC 2663.

3. RESULTS

Sequence data for ITS, LSU and EF1a genes were obtained for isolates *Phaeosaccardinula multiseptata* (IFRDCC 2639), *P. dendrocalami* (IFRDCC 2649, IFRDCC 2663) and *P. ficus* (MFLUCC 10-0009). RPB2 sequence data were only obtained successfully for isolates of

P. dendrocalami. Because ITS and LSU sequence data is only available for most members of *Chaetothyriaceae* in GenBank, we use these genes in the phylogenetic analysis. Individual ITS, LSU, EF1a and RPB2 gene sequences are compared manually for the four *Phaeosaccardinula* strains.

3.1 Phylogeny based on combined LSU and ITS gene data set

The combined LSU and ITS data set utilized 37 taxa with *Venturia inaequalis* as the outgroup taxon. The dataset consists with 14.65% missing and gap characters out of a total set of 1,450 characters and the best scoring tree selected with a final ML Optimization Likelihood -10039.136870. Phylogenetic trees obtained from Bayesian and RAxML analyses yielded a best scoring tree with similar overall topology at the genus and family relationships are in agreement with previous work based on a RAxML tree (Figure 1). The phylogenetic hypothesis strongly supports four monophyletic groups, the three new sequences described herein cluster within the *Phaeosaccardinula* (*Chaetothyriaceae*) clade; they are unrelated to species of *Herpotrichiellaceae*, *Verrucariaceae*, *Trichomeriaceae* and *Capnodiaceae*. The placement of sequence data from strains *Phaeosaccardinula multiseptata*, *P. dendrocalami* and *P. ficus* in this lineage received 100% bootstrap support and Bayesian posterior probabilities of 1.00, while *Phaeosaccardinula multiseptata* clusters in a subclade with *P. ficus* with 100% bootstrap support and Bayesian posterior probabilities of 1.00. The two strains of *P. dendrocalami* cluster in a subclade with 100% bootstrap support and Bayesian posterior probabilities of 1.00.

Comparisons of individual gene data show that *P. multiseptata* and *P. ficus* have four ITS, six LSU and one EF1a base pair difference. The two strains of *P. dendrocalami* have one ITS, one LSU, four EF1a and 13 RPB2 base pairs difference. The further supports *P. multiseptata* being a different species to *P. ficus* while the two strains of *P. dendrocalami* are same species based on molecular evidence.

3.2 Taxonomy

Phaeosaccardinula multiseptata H. Yang & K.D.Hyde, sp. nov. (Figure 2)
MycoBank MB 805931

Etymology: from the Latin *multi* and *septate* in reference to the large number of septa found in the ascospores

Foliar epiphytes growing on the upper surface of living leaves forming soot-like coatings. Mycelium superficial, brown to dark brown, hyphae-like dark brown, reticulate to branched, constricted at the septa. Sexual state: Ascomata 156-182 μm diam, 123-135 μm high ($\bar{x} = 172 \times 124 \mu\text{m}$, $n=10$) orbicular, with depressed central part, black, scattered on surface of leaving leaves, subglobose to globose, lacking setae, without ostiole in mature ascomata. Peridium 26-31 μm wide ($\bar{x} = 28 \mu\text{m}$, $n=20$), thick-walled, inwardly hyaline to dark brown, comprised of cells of *textura prismatica*, brown towards the outside, comprised of cells of *textura angularis*. Ascii 55-69 \times 30-40 μm ($\bar{x} = 64 \times 33 \mu\text{m}$, $n=20$) 4-6-spored, bitunicate, obovoid to oval. Ascospores 38-47 \times 13-16 μm ($\bar{x} = 45 \times 14 \mu\text{m}$, $n=30$), pale brown to brown, oblong-ellipsoid to reniform, muriform, with obscure 9-12 transversal septa and 7-11 longitudinal septa, surface rough, slightly constricted at the septa. Asexual state: Unknown.

Culture characteristics: Colonies on PDA growing slowly, 3 cm diam after 40 days, surface gray to olivaceous gray, spreading, with obvious folds, velvety, water droplets forming on the surface, some dark mycelium branching at the margin. No asexual state produced on PDA after 60 days.

Material examined: CHINA – Yunnan Province, Jinghong, on living leaf of *Dillenia pentagyna* Roxb., 3 December 2011, Hui Yang, BN04 (IFRD 9041,

holotype), ex-type living culture = IFRDCC 2639 (duplicates in MFLUCC and ICMP).

Sequence data: ITS = KF667241, LSU = KF667244, EF1a = KF722789.

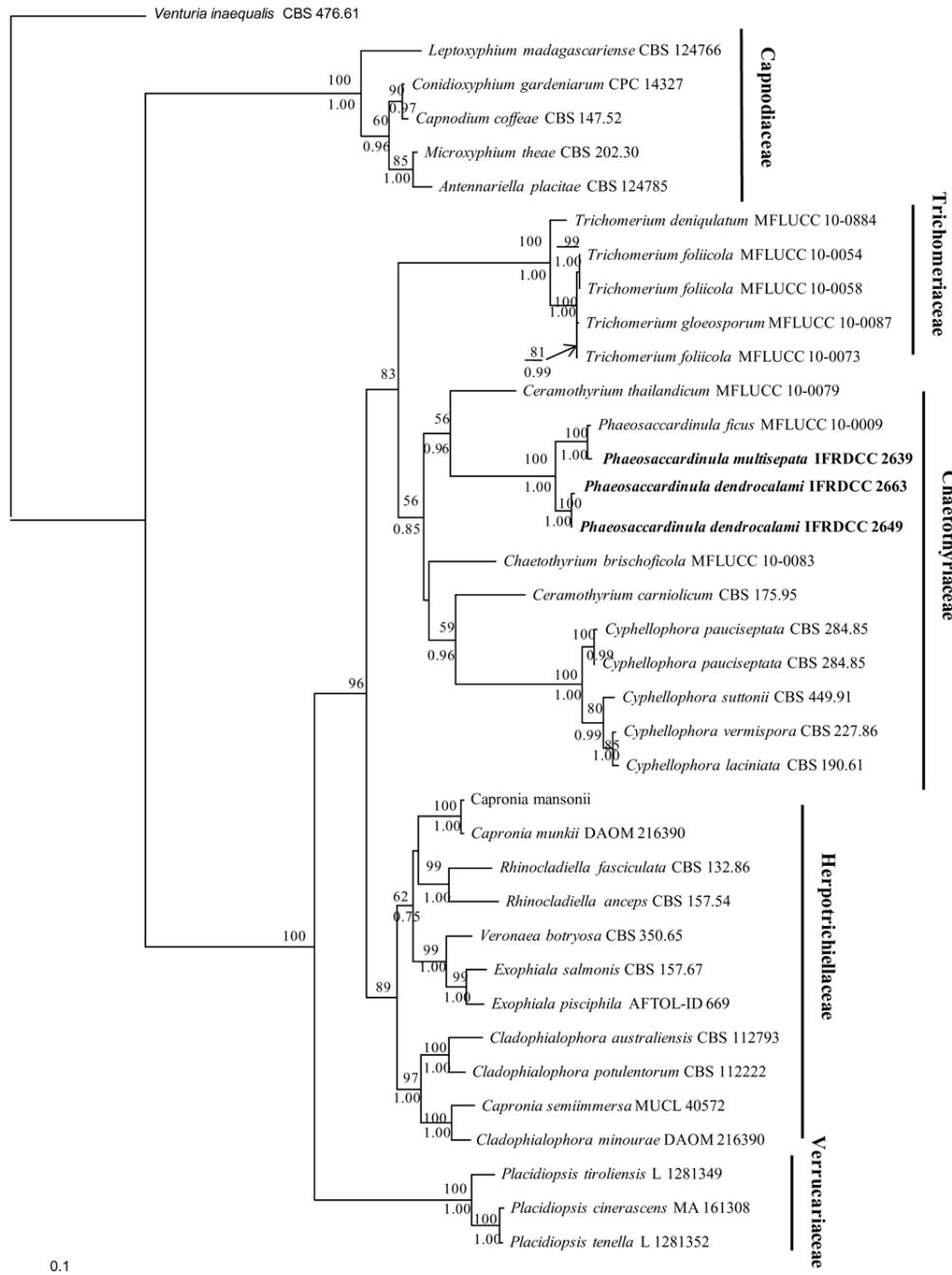


Figure 1. Gene tree constructed using RAxML based on phylogenetic analysis of the nucleotide sequences of combined ITS and 28S nrDNA. Bootstrap support values $\geq 50\%$ and Bayesian values $\geq 80\%$ are shown above or below the branch. *Venturia inaequalis* is the outgroup taxon. The original strain numbers are noted after the species names and names of isolates with newly obtained sequences from this study are in bold.

Notes: This species is similar to *Phaeosaccardinula ficus* which was described from *Ficus* in northern Thailand by Chomnunti *et al.* (2012). Our collection differs from *P. ficus* in having smaller ascomata ($156\text{-}182 \times 123\text{-}135 \mu\text{m}$, $\bar{x}=172 \times 124 \mu\text{m}$, versus $188\text{-}226 \times 116\text{-}193 \mu\text{m}$, $\bar{x}=209 \times 174 \mu\text{m}$) and ascii ($55 \times 69 \times 30\text{-}40$

μm , $\bar{x}=64 \times 33 \mu\text{m}$ vs $120\text{-}185 \times 49\text{-}64 \mu\text{m}$, $\bar{x}=148 \times 55 \mu\text{m}$), and pale brown ascospores, with a large number of transverse (9-12 vs 7-8) and longitudinal (7-11 vs 6-8) septa. The host *Dillenia pentagyna Roxb* also differs. The ITS sequence data have 4 base pair different, LSU have 6 and EF1a have 1 between our collection and *Phaeosaccardinula ficus*.



Figure 2. *Phaeosaccardinula multiseptata* (holotype). A. Sooty mold-like appearance on leaf surface. B. Appearance of colony and ascomata on the host surface. C, D. Vertical sections of ascoma. E, F. Ascii. G-I. Ascospores. Scale bars: C, D = 40 μm , E, F = 10 μm , G-I = 20 μm .

Phaeosaccardinula dendrocalami H. Yang & K.D. Hyde, sp. nov. (Figure 3)
MycoBank MB 805932
Etymology: in reference to its occurrence on *Dendrocalamus*.

Foliar epiphytes growing on the upper surface of living leaves forming a soot-like coating. Mycelium superficial, black, hyphae-like, dark brown to black, reticulate-branched, constricted at the

septa. Sexual state: Ascomata 144-168 μm diam, 105-115 μm high ($\bar{x} = 161 \times 107 \mu\text{m}$, $n = 10$), black, subglobose to globose, lacking setae, with a central ostiole in mature ascomata. Peridium 32-36 μm wide ($\bar{x} = 34 \mu\text{m}$, $n = 20$), thick-walled, inwardly of hyaline of *textura prismatica*, dark brown to brown towards the outside, comprised 3-4 layers of *textura angularis*. Ascii 57-70 \times 27-41 μm ($\bar{x} = 63 \times 32 \mu\text{m}$, $n = 20$) 4-6-spored, bitunicate, oblong-ellipsoid to reniform when young, subglobose to oval when mature, with short pedicel, ocular chamber not visible in mature ascii. Ascospores 37-49 \times 11-15 μm ($\bar{x} = 40 \times 13 \mu\text{m}$,

$n = 30$), hyaline, oblong-ellipsoid to reniform and fusiform, muriform, with 5-7 transversal septa and 3-5 longitudinal septa, guttulate, constricted at the septum, narrow at the ends. Ascospores germinating on PDA within 12 hours, germ tubes developing from numerous cells of ascospores. Asexual state: Unknown.

Culture characteristics: Colonies on PDA growing slowly, 2 cm diam after 40 days, surface gray to olivaceous green, spreading, velvety, dense floss on the surface, pale brown and smooth at the margin. No asexual state produced on PDA after 60 days.

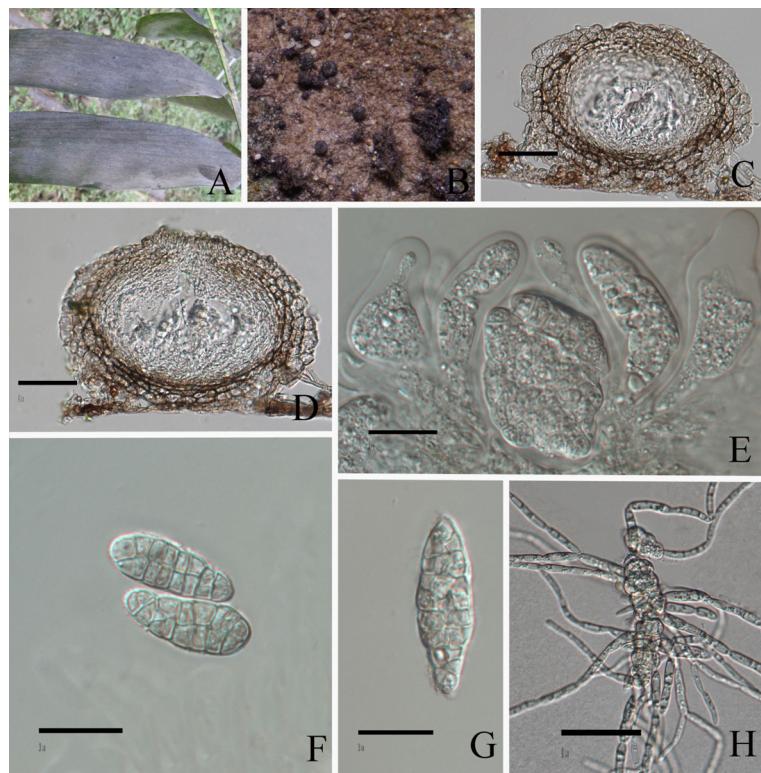


Figure 3. *Phaeosaccardinula dendrocalami* (holotype). A. Sooty mold-like appearance on leaf surfaces. B. Appearance of colony and ascomata on the host surface. C, D. Vertical sections of ascoma. E. Ascii. F. G. Ascospores H. Germinating ascospore. Scale bars: C, D = 40 μm , E-G = 20 μm , H = 40 μm .

Material examined: CHINA – Yunnan Province, Lincang, on living leaf of *Dendrocalamus brandisii*, 17 December 2012, Hui Yang, LC11 (IFRD 9042, holotype), ex-type living culture = IFRDCC 2649. Ibid., Ruili, on living leaf of *Schima wallichii* (DC.) Choisy, 15 December 2012, Hui Yang, RL02 (IFRD 409-001), living culture = IFRDCC 2663 (duplicates in MFLUCC and ICMP).

Sequence data: IFRDCC 2649: ITS = KF667242, LSU = KF667245, EF1a = KF722790, RPB2 = KF 753240

IFRDCC 2663: ITS = KF667243, LSU = KF667246, EF1a = KF722791, RPB2 = KF 753241

Notes: *Phaeosaccardinula dendrocalami* was found on *Dendrocalamus brandisii* (bamboo) and is most similar to *P. ficus* and *P. dictyospora* (Petr.) O. Erikss. *P. ficus* differs from *P. dendrocalami* by its lighter mycelium (brownish and pale brown versus dark brown to black), thinner peridium (20-32 µm, $\bar{x} = 27$ µm versus 32-36 µm, $\bar{x} = 34$ µm), larger ascocata (188-226 × 116-193 µm, $\bar{x} = 209 \times 174$ µm versus 144-168 × 105-115 µm, $\bar{x} = 161 \times 107$ µm) and larger asci (120-185 × 49-64 µm, $\bar{x} = 148 \times 55$ µm vs 57-70 × 27-41 µm, $\bar{x} = 63 \times 32$ µm). *P. dictyospora* differs from *P. dendrocalami* in having light brown, larger ascospores (36-61 × 16-24 µm vs 37-49 × 11-15 µm) and a larger number of transversal septa. The ITS and LSU sequence data have only 1 base pair difference, EF1a have 4 and RPB2 have 13 base pairs difference between strains.

4. DISCUSSION

In this study, we illustrate morphology and provide molecular data from three strains of two collections of *Phaeosaccardinula* (*Chaetothyriaceae*) isolated from Yunnan Province, China. Two new species are

introduced in *Phaeosaccardinula* and illustrated with photomicrographs and descriptions. ITS LSU, EF1a and RPB2 sequence data for the strains are added to GenBank. *Phaeosaccardinula* species are characterized by ascocata that form beneath a mycelial pellicle on the leaf surface, with ascocata and mycelium that lack setae, bitunicate asci and hyaline to light brown, muriform ascospores [6,10]. The phylogenetic analyses of ITS, LSU combined sequences revealed that the three *Phaeosaccardinula* isolates cluster within *Chaetothyriaceae* in the combined analyses of ITS, LSU sequence data and are remote from *Herpotrichiellaceae*, *Verrucariaceae*, *Trichocomeriaceae* and *Capnodiaceae*, and other genera, i.e. *Chaetothyrium*, *Ceramothyrium*, *Cyphellophora* and *Vonarxia* in *Chaetothyriaceae*. The topology and bootstrap support in the RAxML tree and Bayesian posterior probabilities in Figure 1. indicate that all *Phaeosaccardinula* isolates are the very similar or identical as *P. ficus*. However, both *P. dendrocalami* and *P. multiseptata* differ in morphology and are introduced as new species based on morphological differences. Manual comparison of ITS, LSU EF1a and RPB2 sequence data of *P. multiseptata* and *P. ficus* show that the ITS partial gene has four, LSU six and EF1a one base pair difference; ITS has 1, LSU has 1, EF1a has 4 and RPB2 has 13 base pairs differences between the two strains (IFRDCC2649 and IFRDCC2663) of *P. dendrocalami*. These strains were isolated from different hosts so 13 different base pairs differences, may indicate they are also distinct species.

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